Matrix Science Mathematic (MSMK)

DOI: http://doi.org/10.26480/msmk.01.2020.10.13



RESEARCH ARTICLE



ANALYTICAL APPROXIMATION FOR THE NONLINEAR DYNAMICS OF ERK ACTIVATION IN THE PRESENCE OF COMPETITIVE INHIBITOR

Yudi Ari Adia*, M. Irawan Jayadia, Agung Budiantorob

^a Department of Mathematics, Faculty of Applied Science and Technology, Universitas Ahmad Dahlan, Yogyakarta 55191, Indonesia ^b Department of Biology, Faculty of Applied Science and Technology, Universitas Ahmad Dahlan, Yogyakarta 55191, Indonesia *Corresponding Author email: yudiari@uad.ac.id

This is an open access article distributed under the Creative Commons Attribution License CC BY 4.0, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

ARTICLE DETAILS	ABSTRACT
Article History: Received 10 May 2020 Accepted 14 June 2020 Available online 13 July 2020	The extracellular signal-regulated protein kinase (ERK), a subfamily of Mitogen-Activated Protein Kinase (MAPK) pathways, is one of the most important signals in the regulation of many biological processes. Deregulated of MAPK signaling pathways has been observed in human cancers with potential involvement in most of all cellular processes leading to tumorigenesis so that it became a potential target for therapy in cancer patients. In this paper, we discuss a Mathematical model of ERK activation in the presence of a small molecule inhibitor that competes with RAS. We present analytical expressions for the concentration of RAS, complex RAS-ERK, complex RAS-Inhibitor, and activated ERK in terms of dimensionless parameters using He's Homotopy Perturbation Method (HPM). The analytical results are compared with numerical simulation and satisfactory agreement is obtained.
	KEYWORDS

ERK activation, Competitif Inhibitor, Homotopy Perturbation Methods, Analytical Solution.

1. INTRODUCTION

MAPK pathways activated by growth and stress signals as well as cytokines by way of cell receptors such as tyrosine kinase receptors (RTKs) and G-protein coupled receptors (GCPR) (Dhillon et al., 2007). The MAPK signaling pathway is vital in regulating many cellular processes, including inflammation, cell stress response, differentiation, division, proliferation, metabolism, and apoptosis. The known major MAPK cascades distinguished by four distinct cascades, including the extracellular signal-related kinases (ERK1/2), Jun amino-terminal kinases (JNK1/2/3), p38-MAPK and ERK5. Activation of these pathways is started by ligand binding to receptor tyrosine kinases (RTK) at the cell surface and via Ras, Raf, MEK (mitogen-activated protein kinase), culminates in the regulation of gene transcription in the nucleus by the last pathway component, extracellular signal-regulated kinase (ERK) (Knight and Irving, 2014).

The RAS/ERK pathway is activated by the adaptor protein Grb2 that recruited to the tyrosine phosphorylated ErbB1 through its Src homology 2 (SH2) domain. Grb2 then brings in the guanyl nucleotide-release protein SOS to the plasma membrane by Grb2 binding to SOS through its Src homology 3 (SH3) domain. The mitogen-activated protein kinase kinase kinase (MKKK) RAF then binds to the activated RAS. This activates RAF, which leads to the phosphorylation of the mitogen-activated protein kinase kinase (MKK) MEK1/2 following activates the mitogen-activated protein kinase (MAPK) ERK1/2 through phosphorylation. This leads to the

activation of a variety of ERK1/2 downstream signaling pathways, including transcription factors, and ultimately to increased proliferation and cell survival (Henson and Gibson, 2010).

The RAS/ERK pathways are frequently deregulated in human cancers as a result of abnormal activation of receptor tyrosine kinases or gain-offunction mutations, mainly in the RAS or RAF genes. The deregulated activation of the RAS/ERK pathway has been reported in many types of carcinomas that affect the skin breasts, pancreas, liver, and the thyroid, respectively (Santarpia et al., 2012; Cseh et al., 2014; Wellbrock and Arozarena, 2016). In this paper, we focus on the RAS/ERK pathway as an application example for the proposed modeling approach. The deregulated activation of the Ras-ERK pathway includes overexpression of cell membrane receptors and ligands, mutations of receptors and signaling proteins, and the sustained ligand-receptor stimuli mediated by autocrine or paracrine means. This understanding of the RAS-ERK signaling pathway inspired various therapeutic approaches focused on drug-mediated inhibition of the mutant RAS-RAF axis, respected to be the key driver of the deregulated activation of ERK. Unfortunately, this target inhibition is deflected by cancerous cells through the activation of the PI3K-mTOR pathway, whose deregulated control of cell growth and survival coexist with that of the Ras-ERK signaling pathway for many of cancers (Derbal, 2014).

Mathematical modeling of biopathways with molecule inhibition have

Quick Response Code	Access this article online		
	Website: www.matrixsmathematic.com	DOI: 10.26480/msmk.01.2020.10.13	

studied (Adi et al., 2016). The other model of the MAPK pathway also has been conducted in (Derbal, 2014; Pappalardo et al., 2016). In this paper, we develop a Mathematical model of ERK activation in the presence of a small molecule inhibitor that competes with RAS. Several papers have studied the modeling of some nonlinear problems in enzyme kinetics and provide an analytical solution by using the Homotopy Perturbation Method (HPM) (Varadharajan and Rajendran, 2011; Sivasamy et al., 2016). Recently presented the use of HPM to give an analytical approximate solution of the nonlinear problem (Haq, 2019). Now, HPM has been widely used to solving numerous problems of the nonlinear system because of its rapid convergence (Sinan, 2020). In this paper, we use the HPM methods in ERK activation under the condition that the initial ERK concentration is much smaller than the initial concentration of the RAS, $s_0 \ll e_0$. The purpose of this works is to present analytical expressions for the concentration of ERK, complex RAS-ERK, complex RAS-Inhibitor, and activated ERK in terms of dimensionless parameters. We also offer a comparative study of the same with numerical simulation

2. MODEL DEVELOPMENT

by using Runge Kutta fourth order (RK).

In this model, we assume that a small molecule inhibitor act as a competitive inhibitor that mimics ERK but doesn't undergo a reaction. We then follow Ingalls in modeling reaction enzyme subtract in presence competitive inhibitor (Ingalls, 2013). The formulation of biochemical reaction considers a reaction where *ERK* as a substrate bind *RAS* as an enzyme to form a complex *RAS*: *ERK*. The Inhibitor is structurally similar to the substrate so that it competes for the active site by developing a dead-end complex. The schematic representation of this reaction follows:

$$RAS + RAF \underset{d_{1}}{\overset{a_{1}}{\leftrightarrow}} RAS : ERK \xrightarrow{k_{1}} RAS + ERKa$$
$$a_{2}$$
$$RAS + I \underset{d}{\leftrightarrow} RAS : I$$

where a_i , d_i and k_i is the rate of association, dissociation, and activation of protein assumed to be constant.

In order to describe the mathematical model of the biochemical reaction above, we use the law of mass action in an ordinary differential equations (ODE) system. Let us define states of the system; s=[ERK]=concentration of ERK, e=[RAS]=the concentration of RAS, c=[RAS:ERK= the concentration of RAS:ERK complex, p=[ERKa]=the concentration of activated ERK, i=[I]= the concentration of inhibitor, $c_t = [RAS:I] =$ concentration of RAS-Inhibitor complex.

With the law of mass action and quasi-steady-state assumption, the mathematical model is given by the ODE system

$$\frac{ds}{dt} = d_1 c - a_1 s(e_0 - c - c_1)$$
(1)

$$\frac{dc}{dt} = a_1 s(e_0 - c - c_1) - d_1 c - k_1 c$$
(2)

$$\frac{dp}{dt} = k_1 c \tag{3}$$

$$\frac{dc_{I}}{dt} = a_{2}(e_{0} - c - c_{I})i - d_{2}c_{I}$$
(4)

With initial condition

 $s = s_{o}, c = 0, p = 0, c_{i} = 0$ (5)

By introducing the following parameters

$$\tau = \frac{a_1 e_0 t}{\varepsilon}, u(\tau) = \frac{s}{s_0}, v(\tau) = \frac{c}{e_0}, w(\tau) = \frac{c_1}{e_0}, z(\tau) = \frac{p}{e_0},$$

$$\begin{split} \lambda_{1} &= \frac{d_{1}}{a_{1}s_{0}}, \varepsilon = \frac{e_{0}}{s_{0}}, \lambda_{2} = \frac{\left(d_{1} + k_{1}\right)}{a_{1}s_{0}}, \lambda_{3} = \frac{\varepsilon a_{2}i}{a_{1}}, \\ \lambda_{4} &= \frac{\varepsilon \left(a_{2}i + d_{2}\right)}{a_{1}}, \lambda_{5} = \frac{\varepsilon k_{1}}{a_{1}e_{0}}. \end{split}$$

The system of equations (1)- (4) with initial conditions (5) can be represented in dimensionless form as follows:

$$\frac{du}{d\tau} = \lambda_1 \varepsilon v - \varepsilon u + \varepsilon u v + \varepsilon u w \tag{6}$$

$$\frac{dv}{d\tau} = \varepsilon u - uv - uw - \lambda_2 v \tag{7}$$

$$\frac{dw}{d\tau} = \lambda_3 - \lambda_3 v - \lambda_4 w \tag{8}$$

$$\frac{dz}{d\tau} = \lambda_5 v \tag{9}$$

with initial conditions

$$u(0) = 1, v(0) = 0, w(0) = 0, z(0) = 0$$
(10)

3. ANALYTICAL SOLUTION OF STEADY STATE CONCENTRATION USING HPM

To find the analytical expressions for the concentration of *ERK*, *RAS*: *ERK* complex, *RAS*: *Inhibitor* complex, and *ERKa*, we first construct a homotopy to determine the solution of equation (6)-(8) as follows:

$$(1-p)\left[\frac{du}{d\tau} + \varepsilon u\right] + p\left[\frac{du}{d\tau} + \varepsilon u - \lambda_1 \varepsilon v - \varepsilon u v - \varepsilon u w\right] = 0$$
(11)

$$(1-p)\left[\frac{dv}{d\tau} + \lambda_2 v\right] + p\left[\frac{dv}{d\tau} + \lambda_2 v - u + uv + uw\right] = 0$$
(12)

$$(1-p)\left[\frac{dw}{d\tau} + \lambda_4 w\right] + p\left[\frac{dw}{d\tau} + \lambda_4 w - \lambda_3 + \lambda_3 v\right] = 0$$
(13)

with the initial conditions are as follows:

$$u(0)=1, v(0)=0, w(0)=0 \tag{14}$$

Approximate solutions of (11), (12), and (13) are

$$u = u_0 + pu_1 + p^2 u_2 + p^3 u_3 + \dots$$
(15)

$$v = v_0 + pv_1 + p^2 v_2 + p^3 v_3 + \dots$$
 (16)

$$w = w_0 + pw_1 + p^2 w_2 + p^3 w_3 + \dots$$
(17)

After substituting equations (15), (16), and (17) into equations (11), (12), and (13) respectively, and comparing the coefficient of like powers of p we obtain for the ERK concentration:

$$p^0: \frac{du_0}{d\tau} + \varepsilon u_0 = 0 \tag{18}$$

$$p': \frac{du_1}{d\tau} + \varepsilon u_1 - \varepsilon \lambda v_0 - \varepsilon u_0 v_0 - \varepsilon u_0 w_0 = 0$$
⁽¹⁹⁾

$$p^{2}:\frac{dv_{2}}{d\tau}+\varepsilon u_{2}-\varepsilon \lambda_{1}v_{1}-\varepsilon u_{0}v_{1}-\varepsilon u_{1}v_{0}$$
(20)

 $-\alpha u_{_0}w_{_1} - \alpha u_{_1}w_{_0} = 0$ For *RAS: ERK* complex concentration we have

$$p^{\circ}: \frac{dv_{\circ}}{d\tau} + \lambda_2 v_{\circ} = 0$$

$$p^{1}:\frac{dv_{1}}{d\tau} + \lambda_{2}v_{1} - u_{0} + u_{0}v_{0} - u_{0}w_{0} = 0$$
(22)

(21)

(23)

$$p^{2}: \frac{dv_{2}}{d\tau} + \lambda_{2}v_{2} - u_{1} + u_{1}v_{0} + u_{0}v_{1} + u_{0}w_{1} - u_{1}w_{0} = 0$$

For RAS: Inhibitor complex concentration we have:

$$p^{0}: \frac{dw_{0}}{d\tau} + \lambda_{4}w_{0} = 0$$
⁽²⁴⁾

$$p': \frac{dw_1}{d\tau} + \lambda_4 w_1 - \lambda_3 + \lambda_3 v_0 = 0$$
⁽²⁵⁾

$$p^{2}: \frac{dw_{2}}{d\tau} + \lambda_{4}w_{2} + \lambda_{3}v_{1} = 0$$
⁽²⁶⁾

Solving (18)-(26) using initial conditions (14) and according to HPM, we set $p \rightarrow 1$ in (15)-(17) and obtain the final results for solutions (6)-(10) as follows:

$$u(\tau) = e^{-\varepsilon\tau} + \frac{\varepsilon\lambda_{1}\tau e^{-\varepsilon\tau}}{\lambda_{2} - \varepsilon} + \frac{\lambda_{1}\varepsilon\left(e^{-\lambda_{2}\tau} - e^{-\varepsilon\tau}\right)}{\left(\lambda_{2} - \varepsilon\right)^{2}} - \frac{\left(e^{-\varepsilon\tau} - e^{-2\varepsilon\tau}\right)}{\left(\lambda_{2} - \varepsilon\right)} - \frac{\varepsilon\left(e^{-(\varepsilon+\lambda_{2})\tau} - e^{-\varepsilon\tau}\right)}{\lambda_{2}\left(\lambda_{2} - \varepsilon\right)} + \frac{\varepsilon\lambda_{3}\tau e^{-\varepsilon\tau}}{\lambda_{4}^{2}\left(\lambda_{2} - \varepsilon\right)} - \frac{\varepsilon\lambda_{3}e^{-\varepsilon\tau}}{\lambda_{4}^{2}},$$

$$(27)$$

$$v(\tau) = \frac{\left(e^{-\varepsilon\tau} - e^{-\lambda_{2}\tau}\right)}{\lambda_{2} - \varepsilon} + \frac{\left(e^{-\lambda_{2}\tau} - e^{-(\varepsilon + \lambda_{2})\tau}\right)}{\varepsilon(\lambda_{2} - \varepsilon)}$$

$$-\frac{\left(e^{-2\varepsilon\tau} - e^{-\lambda_{2}\tau}\right)}{\left(\lambda_{2} - \varepsilon\right)\left(\lambda_{2} - 2\varepsilon\right)}$$

$$+ \frac{\lambda_{3}\left(e^{-(\varepsilon + \lambda_{4})\tau} - e^{-\lambda_{2}\tau}\right)}{\lambda_{4}\left(\lambda_{2} - \varepsilon - \lambda_{4}\right)} + \frac{\lambda_{3}\left(e^{-\lambda_{2}\tau} - e^{-\varepsilon}\right)}{\lambda_{4}\left(\lambda_{2} - \varepsilon\right)}$$
(28)

$$w(\tau) = \frac{\lambda_3}{\lambda_4} \left(1 - e^{-\lambda_4 \tau} \right) + \frac{\lambda_3 \left(e^{-2\lambda_4 t} - e^{-\lambda_2 \tau} \right)}{\left(\lambda_4 - \varepsilon \right) \left(\lambda_2 - \varepsilon \right)} - \frac{\lambda_3 \left(e^{(\lambda_4 - \varepsilon)\tau} + e^{-\lambda_4 \tau} \right)}{\left(\lambda_4 - \varepsilon \right) \left(\lambda_4 - \lambda_2 \right)}$$
(29)

The dimensionless concentration of active ERK, ERKa is given by

$$z(\tau) = \lambda_{5} \int_{0}^{\tau} v(\tau) d\tau$$
(30)

Equatian (30) is easy to solve, represent the product $z(\tau)$ for all values parameters given in equation (28).

4. RESULT AND DISCUSSION

The concentration of *RAS*, complex *RAS* – *RAF*, complex *RAF*: Inhibitor, and Activated ERK versus time is plotted in Figure 2 using (27) - (30). The nonlinear ODE system (1) - (4) with initial condition (5) is also solved using numerical methods. The numerical method is done by using the Runge-Kutta fourth order with Ode45, one of differential solvers in Matlab. Figure 1 shows the numerical result, and we have a good agreement with analytical expression using HPM in Figure 2. It is observed that the dimensionless *RAS* concentration, *u* and complex *RAS*: *RAF*, *v* decreases gradually from its initial value and becomes zero. It is also observed that the concentration ERK, *z* and complex *RAS*: *I* w increases. This simulation result alsoggests that ERK is a potential target for improving the success of molecular therapy. We also give comparation these two methods in Figures 3 - 6 for various values of dimensionless parameters \mathcal{E} .



Figure 1: Plot of dimensionless concentration of ERK, complex RAS-ERK, complex RAS-Inhibitor, and ERKa numerically using Runge Kutta fourth-order (RK) parameter value $\varepsilon = 10$, $\lambda_1 = 0.5$, $\lambda_2 = 2$, $\lambda_3 = 0.2$, $\lambda_4 = 0.7$,







Figure 3: The concentration of dimensionless ERK (*u*) using HPM $(-\Delta -)$ and RK (--) for various values of dimensionless parameters \mathcal{E} .



Figure 4: The concentration of dimensionless complex *RAS*: *ERK* (*v*) using HPM $(-\Delta -)$ and RK (--) for various values of dimensionless parameters \mathcal{E} .





parameters \mathcal{E} .



Figure 6: The concentration of dimensionless ERKa (z) using HPM ($-\Delta$ -) and RK (--) for various values of dimensionless parameters \mathcal{E} .

From Figures 3 – 6, it is infered that if the initial concentration of ERK is getting smaller than the contentration of RAS, indicated with the increase value of the dimensionless parameter ε , HPM provide rapid and fast convergence.

5. CONCLUSION

In this paper, an analytical expression of ERK, complex *RAS*: *ERK*, complex *RAS*: Inhibitor and *ERKa* in terms of dimensionless parameters are derived using He's Homotopy Perturbation Method (HPM). The HPM compared with numerical solutions method, i.e. Runge Kutta fourth-order. We compared both types of solutions graphically and observed that the solution gets closer to each other. This result suggests that the HPM gives a very accurate approximation to the solution of our model under the assumption that the initial concentration of RAS is higher than the initial concentration of ERK.

REFERENCES

- Adi, Y.A., Adi-Kusumo, F., Aryati, L., Hardianti, M.S., 2016. Modelling Inhibition of AKT Phosphorylation in Acute Myeloid Leukemia. AIP Conference Proceedings, Pp. 1746. doi:10.1063/1.4953987
- Cseh, B., Doma, E., Baccarini, M., 2014. "RAF" neighborhood: protein interaction in the Raf/Mek/Erk pathway. Febs Letters, Pp. 2396-2406.
- Derbal, Y., 2014. State Machine Modeling of MAPK Signaling Pathways. IEEE Engineering in Medicine and Biology Conference.
- Dhillon, A.S., Hagan, S., Rath, O., Kolch, W., 2007. MAP Kinase Signalling Pathways in Cancer. Oncogene, 26 (22), Pp. 3279-3290.
- Haq, I.U., 2019. Analyitcal Approximate Solution of Non-linear Problem by Homotopy Perturbation Method (HPM). Matrix Science Mathematic (MSMK), 3 (1), Pp. 20- 24.
- Henson, E.S., Gibson, S.B., 2010. Epidermal Growth Factor (EGF) Receptor Signaling and Cancer, A. Sitaramayya (ed.), Signal transduction Pathways, Mechanism and Diseases. Springer-Verlag.
- Ingalls, B., 2013. Mathematical Modelling in System Biology an Introduction. Waterloo USA.
- Knight, T., Irving, J.A.E., 2014. Ras/Raf/MEK/ERK Pathway activated in chilhod acute lymphoblastic leukemia and its therapeutic targeting. frontiers in Oncology, 4 (160).
- Pappalardo, F., Russo, G., Candido, S., Pennisi, M., Cavalleri, S., Motta, S., Mc Cubrey, J.A., Nicolleti, F., Libra, M., 2016. Computation Modellinng of PI3K/AKT and MAPK Signalling Pathways in Melanoma Cancer. Plos One, 11 (3).
- Santarpia, L., Lippman, S.L., El-Nagar, A.K., 2012. Targeting the Mitogen Activated Protein Kinase RAS-RAF Signaling Pathway in Cancer Therapy. Expert Opinion on Therapeutic Targets, 16 (1), Pp. 103 - 119.
- Sinan, M., 2020. Analytic Approximate Solution of Rabies Transmission Dynamics Using Homotopy Perturbation Method. Matrix Science Mathematics (MSMK), 4 (1), Pp. 01 - 05.
- Sivasamy P., Ganapathy, J.R.P., Thinakaran I.a, Rajendran, L, 2016. "Enzyme Kinetic modelling and analytical solution of nonlinear rate equation in the transformation of D-methionine into L-methionine in batch reactor using the new homotopy perturbation method," Quim Nova, vol. 39, pp. 1184-1191.
- Varadharajan, G., Rajendran, L., 2011. Analytical Solution of Coupled Non-Linear Second Order Reaction Differential Equations in Enzyme Kinetics. Natural Science, 3, Pp. 459-465.
- Wellbrock, C., Arozarena, I., 2016. The Complexity of the ERK/MAPKinase Pathway and the Treatment of Melanoma Skin Cancer. Front Cell Dev Biol., 4 (33).