# Editor/Author Correspondence

## Edito **DELETE**

r Subject: [IJPHS] Editor Decision

2021- The following message is being delivered on behalf of International Journal of Public

04-16 Health Science (IJPHS).

12:51 PM

Dear Prof/Dr/Mr/Mrs: Azis Ikhsanudin,

We have reached a decision regarding your submission entitled "Topical Anti-Inflammatory Activity of Herbal Gel Formulation Indonesian Nutmeg seeds (Myristica fragrans, Houtt.)" to International Journal of Public Health Science (IJPHS), a peer-reviewed and an OPEN ACCESS journal that makes significant contributions to major areas of public health science.

Our decision is to ACCEPT with revisions

The goal of your revised paper is to describe novel technical results.

A high quality paper MUST has:

- (1) a clear statement of the problem the paper is addressing --> explain in "Introduction" section
- (2) the proposed solution(s)/method(s)/approach(es)/framework(s)/ ....
- (3) results achieved. It describes clearly what has been done before on the problem, and what is new.

In preparing your revised paper, you should pay attention to:

1. Please ensure that: all references have been cited in your text; Each citation should be written in the order of appearance in the text; The references must be presented in numbering and CITATION ORDER is SEQUENTIAL [1], [2], [3], [4], ......

Please download & study our published papers for your references:

- http://ijphs.iaescore.com
- http://ijere.iaescore.com
- http://journal.uad.ac.id/index.php/edulearn
- http://iaescore.com/journals (other journals)

(Please use "Search" menu under "JOURNAL CONTENT" menu in right side of the site)

- 2 An Introduction should contain the following three (3) parts:
- Background: Authors have to make clear what the context is. Ideally, authors should give an idea of the state-of-the art of the field the report is about.
- The Problem: If there was no problem, there would be no reason for writing a manuscript, and definitely no reason for reading it. So, please tell readers why they should proceed reading. Experience shows that for this part a few lines are often sufficient.
- The Proposed Solution: Now and only now! authors may outline the contribution of the manuscript. Here authors have to make sure readers point out what are the novel

aspects of authors work. Authors should place the paper in proper context by citing relevant papers. At least, 5 references (recently journal articles) are used in this section.

3. Results and discussion section: The presentation of results should be simple and straightforward in style. This section report the most important findings, including results of statistical analyses as appropriate. You should present the comparison between performance of your approach and other researches. Results given in figures should not be repeated in tables. It is very important to prove that your manuscript has a significant value and not trivial.

Please submit your revised paper within 6 weeks.

I look forward for hearing from you

Thank you

Best Regards, Dr. Lina Handayani Universitas Ahmad Dahlan ijphs@iaescore.com

-----

Update your metadata in our online system when you submit your revised paper through our online system, included:

- Authors name are presented without salutation
- Authors Name are presented Title Case (ex: Michael Lankan, and NOT written--> michael lankan or MICHAEL LANKAN). Add all authors of your paper as per your revised paper
- Title of revised paper (ex: Application of space vector  $\dots$  , NOT --> APPLICATION OF SPACE VECTOR  $\dots$ )
- Your abstract

Reviewer D:

Does the paper contain an original contribution to the field?:

Is the paper technically sound?:

Yes

Yes

Does the title of the paper accurately reflect the major focus contribution of this paper?: Yes

Please suggest change of the title as appropriate within 10 words: the title cannot be longer than 10 words, rewrite

```
Is the abstract a clear description of the paper?
Yes
Please suggest change of the abstract
Abtract cannot be longer than 200 words
what research design does this research use?
Is the paper well written (clear, concise, and well organized)?:
Yes
Are the equations, figures and tables in this journal style, clear, relevant, and are the
captions adequate?:
Yes
Please score the paper on a scale of 0 - 10 as per the directions below:
9-10 Excellent - Outstanding
7-8 Good
5-6 Average
3-4 Poor
0-2 Very Poor
6
Comments to the Authors (how to improve this paper)::
what research design does this research use?
implications for future research?
Reviewer E:
Does the paper contain an original contribution to the field?:
Yes
Is the paper technically sound?:
Yes
```

Does the title of the paper accurately reflect the major focus contribution of this paper?: Yes Please suggest change of the title as appropriate within 10 words: the title is good enough Is the abstract a clear description of the paper? Yes Please suggest change of the abstract Is the paper well written (clear, concise, and well organized)?: Yes Are the equations, figures and tables in this journal style, clear, relevant, and are the captions adequate?: Yes Please score the paper on a scale of 0 - 10 as per the directions below: 9-10 Excellent - Outstanding 7-8 Good 5-6 Average 3-4 Poor 0-2 Very Poor 6 Comments to the Authors (how to improve this paper):: I provide feedback regarding your manuscript in the text directly. However, your paper will be greater if you can improve the discussion part. International Journal of Public Health Science (IJPHS) http://ijphs.iaescore.com Edito **DELETE** Subject: [IJPHS] Editor Decision 2021-04-20

The following message is being delivered on behalf of International Journal of Public Health Science (IJPHS).

\_\_\_\_\_

-- Authors must strictly follow the guidelines for authors at http://iaescore.com/gfa/ijphs.docx

- -- Number of minimum references is 25 sources (mainly journal articles) for research paper
- -- and minimum 50 sources (mainly journal articles) for review paper

-----

-----

Dear Prof/Dr/Mr/Mrs: Azis Ikhsanudin,

It is my great pleasure to inform you that your paper entitled "Topical Anti-Inflammatory Activity of Herbal Gel Formulation Indonesian Nutmeg seeds (Myristica fragrans, Houtt.)" is ACCEPTED and will be published on the International Journal of Public Health Science (IJPHS).

Please prepare your final camera-ready paper (in MS Word or LATEX file format) adheres to every detail of the guide of authors (MS Word: http://iaescore.com/gfa/ijphs.docx, or http://iaescore.com/gfa/ijphs.rar for LATEX file format), and check it for spelling/grammatical mistakes. Then you should upload your final paper though our online system (as "author version" under our decision, NOT as new submission).

You should submit your camera-ready paper along with your payment receipt and similarity report (that less than 25%) within 6 weeks.

I look forward to hearing from you.

Thank you

Best Regards, Dr. Lina Handayani Universitas Ahmad Dahlan ijphs@iaescore.com

-----

-----

Please ensure that all references have been cited in your text. Each citation should be written in the order of appearance in the text in square brackets. For example, the first citation [1], the second citation [2], and the third and fourth citations [3,4]. When citing multiple sources at once, the preferred method is to list each number separately, in its

own brackets, using a comma or dash between numbers, as such: [1], [3], [5] or [4-8]. It is not necessary to mention an author's name, pages used, or date of publication in the intext citation. Instead, refer to the source with a number in a square bracket, e.g. [9], that will then correspond to the full citation in your reference list. Examples of in-text citations:

This theory was first put forward in 1970 [9]."

Bloom [10] has argued that...

Several recent studies [7], [9], [11-15] have suggested that....

...end of the line for my research [16]......

.-----

-----

In order to cover part of the publication cost, each accepted paper is charged: USD 215 (~IDR 3000K). This charge is for the first 8 pages, and if any published manuscript over 8 pages will incur extra charges USD 50 (~IDR 700K) per page

The payment should be made by bank transfer (T/T):

-----

Bank Account name (please be exact)/Beneficiary: LINA HANDAYANI

Bank Name: CIMB NIAGA Bank

Branch Office: Kusumanegara Yogyakarta

City: Yogyakarta Country: Indonesia

Bank Account #: 760164155700 (formerly: 5080104447117)

SWIFT Code: BNIAIDJAXXX (PT. BANK CIMB NIAGA, TBK. in JAKARTA)

\_\_\_\_\_

-----

## IMPORTANT!!!

- You should submit your payment receipt (along with your camera-ready paper along with your similarity report) within 6 weeks to email: ijphs@iaescore.com, cc: linafkm@gmail.com.
- All correspondence should be addressed to the emails (support by phone is not provided).

International Journal of Public Health Science (IJPHS) http://ijphs.iaescore.com

## **International Journal of Public Health Science (IJPHS)**

Vol. x, No. x, March 2020, pp. xx~xx

# **Anti-Inflammatory Activity of Indonesian Nutmeg Seeds : A Topical Gel Formulation**

## Azis Ikhsanudin<sup>1\*</sup>, Lolita Lolita<sup>1,2</sup>, Daniar Dyanas Rais<sup>1</sup>

<sup>1</sup>Faculty of Pharmacy, Universitas Ahmad Dahlan, Yogyakarta Province 55164, Indonesia <sup>3</sup>Research Division of Clinical Pharmacology, the First Affiliated Hospital of Nanjing Medical University, P.R. China

#### **Article Info**

#### Article history:

Received Revised Accepted

## Keywords:

Nutmeg seed Myristicin Herbal gel Anti-inflammatory

#### **ABSTRACT**

Herbal medicines have been shown as anti-inflammatory with potentially lesser side effects. The active compound of nutmeg seed is proven to accelerate the healing process of inflammation. This study aims to evaluate the gel formulation of Indonesian nutmeg seed extract for anti-inflammatory activity. A true experimental post test only with control group design was used in this study. The gel was formulated with various concentrations of nutmeg seed extract, namely formulations F1 (0%), F2 (2%), F3 (4%), F4 (8 %), and F5 (12%). Analysis of variance (ANOVA) followed by the least significant difference (LSD) methods were performed with SPSS version 22. The results showed that all formulas had an opaque physical appearance, brownish-yellow color, soft texture, and aromatic odor. The increase of extract concentration in gel formula will affect the adhesion and spreadability. F5 showed the highest anti-inflammatory activity compared to other groups. This formula was generally identified as having a good physical appearance, homogeneity, and stability with a pH value of  $6.16 \pm 0.24$ , adhesiveness of  $51.12 \pm 0.15$  sec, and a spreadability of  $19.54 \pm 0.12$ cm2. Therefore, Indonesian nutmeg has the potential to be wellacceptable as a candidate for topical anti-inflammatory agents in global health benefits

This is an open access article under the CC BY-SA license.



## Corresponding Author:

apt. Azis Ikhsanudin, M.Sc Faculty of Pharmacy, Ahmad Dahlan University,

Prof Dr Soepomo Warungboto Umbulharjo, Yogyakarta, Indonesia

Email: azis.ikhsanudin@pharm.uad.ac.id

#### 1. INTRODUCTION.

Inflammation is a tissue-response cascade defense mechanism for specific inflammatory stimuli or pathogenic infections. This inflammatory reaction involves different cellular and vascular pathways with specific humoral secretions to prevent damage and restore tissue function [1]. Specific receptor cells could also trigger various pro and anti-inflammatory cytokines such as IL-1b, IL-6, IL-1a, and TNF- $\alpha$ , which modulate neutrophil migration to endothelial cells. Neutrophils will be released into circulation upon encountering multiple danger signals by producing inflammatory cytokines. Moreover, monocytes will differentiate into tissue macrophages by phagocytosis of damaged cells and other invading pathogens [2][3][4].

The drugs most commonly used for alleviating acute and chronic inflammation are non-steroidal antiinflammatory drugs (NSAIDs) and steroids (corticosteroids)[5][6]. These medications are typically used in combination because of their slightly distinct actions in granulation tissue. NSAIDs inhibit the cyclooxygenase

Journal homepage: http://ijphs.iaescore.com

enzymes (COX-1 and COX-2) responsible for regulating many cellular processes during the inflammatory response [7]. Steroids could suppress inflammation intensely by binding to receptors that control phospholipase A2, COX-2, iNOS, and interleukin. Unfortunately, two classes of drugs should be taken with caution because of the potential adverse events, e.g., gastrointestinal bleeding, myocardial infarction, and kidney disorders [8][9][10].

The development of a plant-based drug is currently the primary alternative approach regarding the long-term risk of using anti-inflammatory drugs. Under these circumstances, medicinal plants have significantly rediscovered cellular pathways to provide potent active compounds as therapeutic phytochemicals [11]. Furthermore, the market for herbal medicines accounts for 83% worldwide in the treatment of inflammatory diseases. It is estimated that it will reach a value of approximately more than 95% in the forthcoming years due to increased revenues from these preparations. Indonesia is the largest producer and exporter of nutmeg in the global market. Overall export volume tended to increase by an average of 3.07% per year during 2009-2018 period [12]. Indonesia nutmeg is of good quality due to its low and almost non-existent aflatoxin content [13]. The myristin content in Indonesian nutmeg was 8.72% higher than Indian nutmeg, which was only 3.8% [14], [15].

A feasibility study reports that myristicin has been recognized in pharmacological mechanisms as a potent anti-inflammatory activity. Several studies have shown that nutmeg seed essential oil containing myristicin and aluminometasilicate was useful for anti-inflammatory activity. It works by inhibiting the biosynthesis of prostaglandin in the human colon. The high myristicin content in nutmeg could play a role in reducing TNF- $\alpha$  levels. Myristicin also demonstrates a similar mechanism as NSAIDs such as indomethacin and aspirin [16][17][18][19].

Topical drug delivery systems are gaining popularity in developing local and systemic drug delivery systems. This system could avoid gastrointestinal irritation, overcome the "first pass" effect and maximize drug concentrations at receptor site [20]. Furthermore, the gel has biodegradable, biocompatible, consistent properties, reasonable penetration rates, and longer retention time [21][22][23]. Previous studies have reported the topical gel formulation of ethanol extract of nutmeg using the albumin denaturation method. This finding showed that nutmeg extract gel could inhibit albumin's denaturation higher than the marketed synthetic drug gels [24]. In our recent study, we used the carrageenan-induced rat paw edema model to evaluate anti-inflammatory activity. The measurement of anti-inflammatory activity was based on the increase in the volume of rat leg edema, and the percentage was calculated based on the AUC value. This approach is more representative for determining the actual anti-inflammatory activity for topical drug preparations. Therefore, our study aimed to design and evaluate Indonesian nutmeg seed extract's topical anti-inflammatory activity using the carrageenan-induced rat paw edema model. This study's significance is expected to produce potent plant-based anti-inflammatory properties based on good efficacy and acceptance standards.

## 2. RESEARCH METHOD

The research method used true experimental post test only with control group. This study aimed to compare the anti-inflammatory activity between formula group of topical herbal gel from Indonesian nutmeg seed extract.

## 2.1 Plant Materials and Chemical Reagents

We collected fresh nutmeg seeds from Malikrubu Regency, Ternate, North Maluku, Indonesia. The plant was identified and approved by the Biology Laboratory of the Faculty of Mathematics and Natural Sciences, Ahmad Dahlan University. The chemical reagents such as distilled water, carbopol 940, methylparaben, propylene glycol, and triethanolamine were obtained through Brataco, Ltd, Indonesia. Voltaren gel containing 1% diclofenac sodium was obtained from the community pharmacist store, Ahmad Dahlan University. All of the reagents were of analytical and pharmaceutical grade.

## 2.2 The Extraction of Indonesian Nutmeg Seed

The purpose of the extraction was to preserve myristicin as an anti-inflammatory bioactive compound in nutmeg seed. Sample preparation was accomplished by peeling the fresh nutmeg using sharp stainlesssteel knives. Afterward, nutmeg seeds should be washed and dried in an oven at  $40^{\circ}$ C for 48 hours. The dried nutmeg seed should be ground to 0.4mm in size using an electric blender to enhance the contact surface. The seed extract was prepared by maceration using 70% ethanol as a solvent for three days at room temperature. About 500 g of nutmeg seed powdered was loaded using 1 liter of 70% ethanol for maceration, followed by stirring with a magnetic stirrer for 2 hours. The supernatant liquid mixture was poured onto filter paper and allowed to stand for 24 hours before being filtered. The whole liquid was filtered using a Buchner porcelain funnel. This treatment procedure was repeated for 3 x 24 hours to obtain a clear filtrate. Further, a gradual evaporation process was performed to obtain a concentrated extract [25].

## 2.3 Topical Gel Formulation of Indonesian Nutmeg Seed Extract

The gels were formulated using different concentration of nutmeg seeds extract ethanol (0%; 2%; 4%; 8% and 12%), carbopol 940 (1.5%), ethanol 96% (4%), propylene glycol (10%), glycerin (20%), triethanolamine (0.8%), methylparaben (0.1%), polysorbate 80 (0.8%), and purified water (q.s to 100%). We prepared the gelbased mixture by mixing carbopol in purified water and maintained it with magnetic stirring until homogeneous (mixture A). Propylene glycol and triethanolamine are dissolved in mixture A to obtain a swollen gel with a pH of 5-6 (mixture B). Afterward, add the nutmeg ethanol extract to mixture B and stir with methylparaben to form a stable and homogeneous gel. Furthermore, the nutmeg extract gels must be stored in a tightly-closed container for 24 hours until the bubbles are removed [24]. The ethanol extract gel formula of nutmeg seed was shown in Table 1

Table 1. Gel formula of ethanol extract nutmeg seed

Ingredient		Formula (gram)							
	F1	F2	F3	F4	F5				
Nutmeg seed extract	0	1	2	4	6				
Carbopol 940	0.75	0.75	0.75	0.75	0.75				
Ethanol 96%	2	2	2	2	2				
Propylenglycol	5	5	5	5	5				
Glyserin	10	10	10	10	10				
Trietanolamin	0.4	0.4	0.4	0.4	0.4				
Methyl paraben	0.05	0.05	0.05	0.05	0.05				
Polysorbate 80	0.4	0.4	0.4	0.4	0.4				
Purified water ad	50	50	50	50	50				

Note: The positive control (F6) used voltaren 50 g gel containing 1% diclofenac sodium

## 2.4 Evaluation of Formulated Nutmeg Seed Gel

The formulated nutmeg seed gel was characterized by specific physical properties such as organoleptic, pH, spreadability, and adhesivity. This parameter has the function of assessing the quality of the gel in various formulas. Another parameter was to investigate the potential activity of the gel as an anti-inflammatory. The test procedure was performed using the carrageenan-induced rat paw edema model. This acute inflammation model is well-accepted and has long been used to determine its potent anti-inflammatory effect.

## 2.4.1. Organoleptic properties

Gel formulations with or without nutmeg seed extract were evaluated organoleptically for color, texture, and homogeneity. These characteristics were performed by visual observation. The formulation consistency and the particle's coarseness were checked by texture and homogeneity parameters. To assess these parameters, we pressed a small amount of formulated gel between the thumb and the index finger.

#### 2.4.2 pH Measurement

Electrochemical pH measurement is used to determine the acidity or alkalinity through a digital pH meter (HANNA HI9813-6 Portable). The principle step was to weigh 1 gram of each gel formula and completely immersed it in the glass electrode. All measurements were taken three times, for which the average reading was recorded [26].

## 2.4.3 Spreadability

The prepared gel spreadability was intended to evaluate the gel's ability to spread and absorb on the skin. This technique was performed by measuring the diameter of spread of 1 gram of gel samples on two horizontal glass slides (10x20cm2) after 60 sec. The standard upper plate for determining gel spreadability was 0.5 g. Each gel formulation was recorded in triplicate [24].

## 2.4.4 Adhesivity

The purpose of gel adhesion testing was to determine the strength that separates the gel from the surface. A tensile adhesion test measures this by placing 2 grams of gel between two resin plates in the designated area. Then clamp the load of 1 kg on the glass plate for 5 minutes. The static resin plate was attached to the adhesion apparatus by applying a trigger load of 80 grams. Further, the evaluation was carried out to determine the length of time using a stopwatch until the resin plate runs out [27].

## 2.4.5 In-vitro anti-inflammatory activity of nutmeg seed extract

The study protocol has obtained ethical approval from Ahmad Dahlan University Ethics Committee with reference number 011502014. The anti-inflammatory properties of nutmeg seed gel products were measured using carrageenan-induce acute inflammation model in hind leg edema of male Wistar rat. The carrageenan-induced paw edema method is a well-defined model of acute inflammation involving various inflammatory mediators, as described by Winter et al. [28]. Initially, healthy Wistar strains (150-200 grams per each) were obtained from an animal breeding house, Faculty of Veterinary Medicine, Gadjah Mada University, Yogyakarta. Rats kept under laboratory conditions were fed twice daily with standard rat chow and water ad libitum. All rats were randomly allocated into six groups: one negative control group, four intervention groups, and one positive control group. Each group was comprising at least five experimental animals. Before the

experiment, the animal had to fast for 24 hours with access to water. 50  $\mu$ l of 1%  $\lambda$ -carrageenan solution (Sigma-Aldrich, Milan, Italy) was suspended in 1% NaCl solution, which had been prepared 1 hour before the experiment. Furthermore, this solution should be injected into the plantar side of the rat's right hind paw. About 0.2 grams of nutmeg seed gel in a different formula was gently applied 50 times with the index finger after 15 minutes of carrageenan injection. The rat paw edema volume was measured immediately using a plethysmometer (model UGO BASILE S.R.L: 7141) at 15 minutes to 3 hours after carrageenan injection. The percentage of inflammation was calculated by the following formula:

% Inflammatory = 
$$\frac{\text{V0} - \text{Vt}}{\text{V0}} \times 100\%$$

Note: Vo and Vt are the volumes of rat paw edema from the control group and experimental group.

#### 2.5 Data Analysis

The study results were reported descriptively on several test data such as organoleptic properties, pH, adhesiveness, and spreadability. The statistical analysis was performed using One-Way ANOVA and followed by the LSD-post hoc test. A *p*-value less than 0.05 was considered statistically significant.

## 3. RESULTS AND DISCUSSIONS

We have successfully formulated herbal gel from the ethanol extract of nutmeg seed in various formulas. Our study demonstrated that the obtained nutmeg seed gel has a good physical appearance and an anti-inflammatory activity due to acute edema reduction. The detailed finding regarding physical characteristics and anti-inflammatory activity will be presented in the subsection below.

## 3.1 Organoleptic properties

As shown in Table 2, the nutmeg seed gel has a good cosmetic appearance with a soft texture, opaque, brownish-yellow in color, characteristic aromatic odor, and homogeneous without segregation. In general, the dense pigment of nutmeg seed contains more dyes to produce a brownish-yellow gel. Previous studies have also reported that the nutmeg seed gel's physical properties were brownish yellow, fragrant and homogeneous[27][24]. The ethanolic extract does not dissolve completely in the gel bases due to the differences in solubility level.

Table 2. Organoleptic properties of nutmeg seed extract gels

Formula	Physical	Color	Odor	Texture	Homogenity	Segregation
	appearance					
1	Transparant	Clear white	Typical of base	Soft	Homogenous	No
2	Opaque	Brownish yellow	Typical of nutmeg oil	Soft	Homogenous	No
3	Opaque	Brownish yellow	Typical of nutmeg oil	Soft	Homogenous	No
4	Opaque	Brownish yellow	Typical of nutmeg oil	Soft	Homogenous	No
5	Opaque	Brownish yellow	Typical of nutmeg oil	Soft	Homogenous	No

The appearance of nutmeg seed extract gel with various extract concentrations is shown in Figure 1.



Figure 1. The physical appearance of nutmeg seed extract gel at F2 (2%); F3 (4%); F4 (8%) and F5 (12%) (from left to right)

## 3.2 pH Measurement

Each nutmeg seed extract gel formula shows the following pH value F1(4.69  $\pm$  0.74), F2 (5.38  $\pm$  0.78), F3 (5.82  $\pm$  0.15), F4 (6.33  $\pm$  0.13), F5 (6.16  $\pm$  0.24) and F6 (5.87  $\pm$  0.12) (Table 3). These pH value has met the standard criteria for topical skin administration (pH ranged 4-7). pH value measurement could be related to the safety and efficacy of topical drug preparations. Gel acidity and alkalinity will affect skin irritation, penetration and stability of active compounds into the skin. The more acidic the pH value of gel will increase skin irritation, while the more alkaline it will cause dry skin [29][27]. The statistic analysis using the LSD post hoc test showed a significant difference in pH value between F3 to F5 compared to F1 (p<0.001). These results mean that the pH value will increase with the addition of nutmeg extract concentration in the gel base. Meanwhile, the pH value of F1 (gel base) was not significantly different from F2 (extract 2%) (p=0.121). This defines that both formulas have a similar pH value. As a gelling agent, each end of carbopol's chain has an acidic carboxylic chemical structure when reacting to water. Carbopol 940 is easily ionized during the

neutralization process with the addition of triethanolamine. Therefore, the interaction between carbopol 940 and triethanolamine could affect the pH stability based on the increase in extract concentration [30].

Table 3	The nH	value	of nutmeg	seed	extract or	1

Formula	pH (Mean $\pm$ SD)	<i>p</i> -value (LSD post hoc test)						
		F1	F2	F3	F4	F5		
F1	4.69±0.74	-	0.121	0.020*	0.002*	0.050*		
F2	5.38±0.78	0.121	-	0.306	0.041*	0.084		
F3	5.82±0.15	0.020*	0.306	-	0.234	0.419		
F4	6.33±0.13	0.002*	0.041*	0.236	-	0.680		
F5	6.16±0.24	0.005*	0.084	0.419	0.680	-		

Note: \* p-value less than 0.05

## 3.3 Adhesivity

Penetration of the active compound gel into the skin will increase along with the enhancement of gel adhesivity properties. The results showed that the average adhesion score of nutmeg seed extract gel was F1 (13.57 $\pm$ 0.11), F2 (18.32 $\pm$ 0.29), F3 (44.33 $\pm$ 0.09), F4 (56.54 $\pm$ 0.19), and F5 (51.12 $\pm$ 0.15) (Table 4). Based on a one-way ANOVA test followed by LSD statistical analysis, the nutmeg extract gel of F4 was significantly greater in adhesion than other formulas (p<0.005). Furthermore, the formulation of F5 with 12% nutmeg seed ethanol extract showed a decrease in adhesiveness compared to F4. This decrease might be due to the pH value being smaller at F5 than F4. The cross-linking between carbopol and other molecules, such as solvents, could also result in ionic attraction, and increasing the gel's viscosity. In acid conditions, increasing the gel's viscosity will have an impact on decreasing the adhesive properties.

Table 4. Adhesive properties of the nutmeg seed extract gel

<b>D</b> .	Mean ± SD (second)					<i>p</i> -value
Parameter	F1	F2	F3	F4	F5	, 1
Adhesivity test (sec)	13.57±0.11	18.32±0.29	44.33±0.09	56.54±0.19	51.12±0.15	0.000

## 3.4 Spreadability

As shown in Table 5, the average spreadability of the nutmeg gel formulations showed F1 ( $9.38\pm0.65$ ), F2 ( $11.54\pm0.21$ ), F3 ( $13.39\pm0.06$ ), F4 ( $14.85\pm0.34$ ), and F5 ( $19.54\pm0.12$ ). These findings concluded that increasing the gel extract concentration would affect the broadest spreadability of the gel. The formula F5 (12%) has the greatest spreadability compared to other formulas. The statistical analysis results through one-way ANOVA stated a significant difference in spreadability between all formulas (p<0.005). Carbopol 940, an acrylic polymer gelling agent, could regulate the viscosity in 1-2% concentrations by producing a three-dimensional matrix to form a viscous gel. The spreadability of the gel formula was highly dependent on the viscosity of the carbopol. The less viscous gelling agent will enhance the spreadability of the gel. The swelling gel process occurs due to the solvent penetration, leading to the cross-linked polymer network to maintain the dosage form and the binding drug particles. However, the alkaline environment pH will increase the density of negative charge, resulting in the gel swollen and facilitating the drug's release [21][30]. Furthermore, the addition of nutmeg seed extract to the gel base will also affect the formation of hydrogen bonds in the physical cross-linking, which influences a reduced viscosity [21].

Table 5. Spreadability properties of the nutmeg seed extract gel

<b>D</b> .						
Parameter	F1	F2	F3	F4	F5	<i>p</i> -value
Spreadability test (cm)	9.38±0.65	11.54±0.21	13.39±0.06	14.85±0.34	19.54±0.12	0.000

## 3.5 In-vitro anti-inflammatory activity of nutmeg seed extract gel

The evaluation of the nutmeg seed herbal gel's anti-inflammatory activity was observed within 180 min (Figure 2). During this time, carrageenan will significantly increase TNF- $\alpha$ , IL-1 $\beta$ , PGE2, iNOS, and COX-2 proteins in peripheral leg inflammation. Carrageenan-induced leg edema follows a model of acute inflammation which consists of two phases: first, which was detected after about 1 hour and was called the fast phase, with the release of histamine and serotonin, and the second stage was called the late phase with the mediators (kinins, prostaglandins) released after 2 and 3 hours, respectively [31]. A previous study suggested that intraplantar carrageenan injection will stimulate rat paw edema and release inflammatory mediators within 180 min [32]. This study is in line with our findings that decreased edema volume also occurred at 180 minutes after intraplantar carrageenan administration.

Figure 2 showed that the formulation of nutmeg seed extract gel has anti-inflammatory effects. It can be proven that the percentage volume of edema in each formula (F2 to F6) was lower than F1 (gel-based negative control). The anti-inflammatory activity of F5 (12%) was higher than that of F2 (2%) and F3 (4%). Furthermore, F4 (8%) and F5 (12%) had anti-inflammatory activity similar to F6 (positive control). This anti-inflammatory activity is influenced by the main compound contained in the nutmeg, namely myristicin. The active compound of nutmeg seed extract, myristicin, has a vital function as an anti-inflammatory by inhibiting chemokines, cytokines, nitrous oxide, and double-stranded growth factors RNA (dsRNA), which are stimulated by macrophages through calcium [33]. Other quercetin compounds also have anti-inflammatory activity by inhibiting the secretion of TNF- $\alpha$ , IL-6, IL-1 $\beta$ , and nitric oxide (NO) [34].

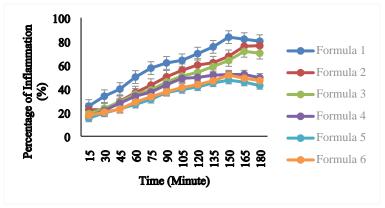


Figure 2. Inflammatory activity of nutmeg seed extract gels.

The volume increase in rat leg edema was used to calculate each formula's anti-inflammatory percentage based on the AUC value. The calculated AUC value and the percentage of anti-inflammatory can be seen in Table 6. Based on the LSD post hoc test analysis reported that the anti-inflammatory activity of F4, F5 did not have a significant difference compared to F6 (positive control). It was concluded that F4 and F5 had similar anti-inflammatory activity as the positive control (1% diclofenac sodium gel in the marketed product). Furthermore, the anti-inflammatory activity of F5 (44.92%) was higher significant difference compared to F4 (37.87%) with p=0.025 The AUC calculation results showed that F5 had the lowest AUC value (5803.72) with the highest percentage of anti-inflammatory activity (44.92%) compared to other formulas (F1 to F5).

Table 6. The percentage of anti-inflammatory activity of nutmeg extract gel was compared between groups.

Formula	AUC	Anti-inflammatory		)				
		activity (%)	F1	F2	F3	F4	F5	F6
F1	10536.94	0	-	0.000*	0.000*	0.000*	0.000*	0.000*
F2	9304.97	11.69	0.000*	-	0.000*	0.000*	0.000*	0.000*
F3	7919.26	24.84	0.000*	0.000*	-	0.000*	0.000*	0.000*
F4	6546.62	37.87	0.000*	0.000*	0.000*	-	0.025*	0.423
F5	5803.727	44.92	0.000*	0.000*	0.000*	0.025*	-	0.124
F6	6295.140	40.25	0.000*	0.000*	0.000*	0.423	0.124	-

Note: \* p-value less than 0.05; F6 is positive control

The highest anti-inflammatory activity of F5 was due to the gel's spreadability and its adhesion to the skin surface. F5 has a good adhesiveness of  $(51.12 \pm 0.15)$  sec with a spreadability of  $(19.54 \pm 0.12)$  cm. The topical drug penetration into the skin was affected by the length of time the gel is in contact with the skin surface. F5 has the highest concentration of nutmeg seed extract (12%) than other formulas (F2 to F4). The high content of myristicin in the F5 gel formula will improve its anti-inflammatory effect by enhancing the drug permeation into the skin. Besides, F5 also had the optimal ternary system composition between ethanol, propylene glycol, and purified water. Propylene glycol (PG) as an enhancer will affect the drug absorption into the skin [35]. A study of topical ibuprofen on human skin reported that ethanol's ternary solvent system, propylene glycol, and water had higher permeability into the skin than the binary solvent system. The ternary system will increase the solubility partition and membrane permeability partitioning by maximizing the active drug flux [36].

## 4. CONCLUSION

These present study indicates that nutmeg gel extract (*Myristica fragrans*, Houtt) with 12% ethanolic extract concentration has a good physical appearance and potent anti-inflammatory activity than other formulas

and marketed product. Further study should implement a novel pharmaceutical technology in topical drug delivery system regarding to enhance the anti-inflammation activity. Hence, Indonesian nutmeg seed extract could be a potential candidate as anti-inflammatory in topical gel dosage form application.

## **ACKNOWLEDGEMENTS**

The author would like to thank the whole researcher in Universitas Ahmad Dahlan who has supported this research.

## REFERENCES

- [1] S. Kany, J. T. Vollrath, and B. Relja, "Cytokines in inflammatory disease," *Int. J. Mol. Sci.*, vol. 20, no. 23, pp. 1–31, 2019, doi: 10.3390/ijms20236008.
- [2] Y. Su, J. Gao, P. Kaur, and Z. Wang, "Neutrophils and macrophages as targets for development of nanotherapeutics in inflammatory diseases," *Pharmaceutics*, vol. 12, no. 12, pp. 1–24, 2020, doi: 10.3390/pharmaceutics12121222.
- [3] A. L. Loktionov, A. I. Konoplya, M. A. Lunev, and A. V. Karaulov, "Immune and oxidant disorders in the pathogenesis of inflammatory periodontal diseases," *Immunologiya*, vol. 36, no. 5, pp. 319–328, 2015, doi: 10.1111/prd.12002.Inflammatory.
- [4] A. Tedgui and Z. Mallat, "Anti-inflammatory mechanisms in the vascular wall," *Circ. Res.*, vol. 88, no. 9, pp. 877–887, 2001, doi: 10.1161/hh0901.090440.
- [5] M. Del Grossi Moura, L. C. Lopes, M. T. Silva, S. Barberato-Filho, R. H. L. Motta, and C. De Cássia Bergamaschi, "Use of steroid and nonsteroidal anti-inflammatories in the treatment of rheumatoid arthritis: Systematic review protocol," *Med. (United States)*, vol. 97, no. 41, 2018, doi: 10.1097/MD.000000000012658.
- [6] A. Lahoti, B. S. Kalra, and U. Tekur, "Evaluation of the analgesic and anti-inflammatory activity of fixed dose combination: non-steroidal anti-inflammatory drugs in experimental animals.," *Indian J. Dent. Res.*, vol. 25, no. 5, pp. 551–554, 2014, doi: 10.4103/0970-9290.147071.
- [7] K. D. Brandt, "The mechanism of action of nonsteroidal antiinflammatory drugs," *J. Rheumatol.*, vol. 18, no. SUPPL. 27, pp. 120–121, 1991, doi: 10.5772/68090.
- [8] P. Kuropakornpong, A. Itharat, S. Panthong, S. Sireeratawong, and B. Ooraikul, "In Vitro and in Vivo Anti-Inflammatory Activities of Benjakul: A Potential Medicinal Product from Thai Traditional Medicine," *Evidence-based Complement. Altern. Med.*, vol. 2020, 2020, doi: 10.1155/2020/9760948.
- [9] K. R. Patil *et al.*, "Animal models of inflammation for screening of anti-inflammatory drugs: Implications for the discovery and development of phytopharmaceuticals," *Int. J. Mol. Sci.*, vol. 20, no. 18, 2019, doi: 10.3390/ijms20184367.
- [10] S. Tsurufuji, K. Sugio, H. Sato, and K. Ohuchi, "A review of mechanism of action of steroid and non-steroid anti-inflammatory drugs," *Inflamm. Mech. Treat.*, no. 202, pp. 63–78, 1980, doi: 10.1007/978-94-010-9423-8 7.
- [11] V. P. Gullo, J. McAlpine, K. S. Lam, D. Baker, and F. Petersen, "Drug discovery from natural products," *J. Ind. Microbiol. Biotechnol.*, vol. 33, no. 7, pp. 523–531, 2006, doi: 10.1007/s10295-006-0107-2.
- [12] H. Juliani Purba, E. Supriadi Yusufi, and J. Hestina, "Performane and Competitiveness of Indonesian Nutmeg in Export Market," *E3S Web Conf.*, vol. 232, p. 02018, 2021, doi: 10.1051/e3sconf/202123202018.
- [13] J. Marmis, N. S. Tongkeles, Lady Lengkey, and S. Ratulangi, "the Quality of Nutmeg Seeds and Mace," vol. 2, pp. 111–121, 2015.
- [14] K. Karmanah, S. Susanto, W. D. Widodo, and E. Santosa, "The Fruit Characteristics of Ambon Forest Nutmeg (Myristica fatua Houtt) and Banda Nutmeg (Myristica fragrans Houtt)," *J. Ilmu Pertan. Indones.*, vol. 25, no. 2, pp. 292–300, 2020, doi: 10.18343/jipi.25.2.292.
- [15] I. P. S. Kapoor, B. Singh, G. Singh, C. S. De Heluani, M. P. De Lampasona, and C. A. N. Catalan, "Chemical composition and antioxidant activity of essential oil and oleoresins of nutmeg (Myristica fragrans Houtt.) fruits," *Int. J. Food Prop.*, vol. 16, no. 5, pp. 1059–1070, 2013, doi: 10.1080/10942912.2011.576357.
- [16] I. Matulyte *et al.*, "The essential oil and hydrolats from myristica fragrans seeds with magnesium aluminometasilicate as excipient: Antioxidant, antibacterial, and anti-inflammatory activity," *Foods*, vol. 9, no. 1, 2020, doi: 10.3390/foods9010037.
- [17] Jinous Asgarpanah, "Phytochemistry and pharmacologic properties of Myristica fragrans Hoyutt.: A review," *African J. Biotechnol.*, vol. 11, no. 65, 2012, doi: 10.5897/ajb12.1043.
- [18] O. A. Olajide, J. M. Makinde, and S. O. Awe, "Evaluation of the pharmacological properties of nutmeg oil in rats and mice," *Pharm. Biol.*, vol. 38, no. 5, pp. 385–390, 2000, doi: 10.1076/phbi.38.5.385.5976.

- [19] W. K. Zhang *et al.*, "Nutmeg oil alleviates chronic inflammatory pain through inhibition of COX-2 expression and substance P release in vivo," *Food Nutr. Res.*, vol. 60, no. April, 2016, doi: 10.3402/fnr.v60.30849.
- [20] M. M. Ghareeb, "Design and in vitro characterization of a topical nanoemulsionenriched hydrogel of econazole nitrate," *J. Appl. Pharm. Sci.*, vol. 9, no. 1, pp. 51–57, 2019, doi: 10.7324/JAPS.2019.90108.
- [21] S. A. A. Nabi, M. A. Sheraz, S. Ahmed, N. Mustaan, and I. Ahmad, "Pharmaceutical Gels: A Review," *RADS J. Pharm. Pharm. Sci.*, vol. 4, no. 1, pp. 40–48, 2016.
- [22] C. Tipa, M. T. Cidade, T. Vieira, J. C. Silva, P. I. P. Soares, and J. P. Borges, "Article a new long-term composite drug delivery system based on thermo-responsive hydrogel and nanoclay," *Nanomaterials*, vol. 11, no. 1, pp. 1–22, 2021, doi: 10.3390/nano11010025.
- [23] R. Sánchez-Sánchez *et al.*, "Development of Hydrogel with Anti-Inflammatory Properties Permissive for the Growth of Human Adipose Mesenchymal Stem Cells," *J. Nanomater.*, vol. 2016, 2016, doi: 10.1155/2016/8654937.
- [24] S. Surendran and A. V. Vidyapeetham, "TOPICAL GEL AND ITS IN VITRO EVALUATION FOR ANTINFLAMMATORY Available Online through Research Article," no. April 2016, 2018.
- [25] E. F. Al-Jumaily and M. H. A. Al-Amiry, "Extraction and Purification of Terpenes from Nutmeg (myristica fragrans)," *J. Al-Nahrain Univ. Sci.*, vol. 15, no. 3, pp. 151–160, 2012, doi: 10.22401/jnus.15.3.21.
- [26] S. Karastogianni, S. Girousi, and S. Sotiropoulos, *pH: Principles and Measurement*, 1st ed., no. December. Elsevier Ltd., 2015.
- [27] R. Aiyalu, A. Govindarjan, and A. Ramasamy, "Formulation and evaluation of topical herbal gel for the treatment of arthritis in animal model," *Brazilian J. Pharm. Sci.*, vol. 52, no. 3, pp. 493–507, 2016, doi: 10.1590/s1984-82502016000300015.
- [28] C. A. Winter, E. A. Risley, and G. W. Nuss, "Carrageenin-Induced Edema in Hind Paw of the Rat as an Assay for Antiinflammatory Drugs," *Proc. Soc. Exp. Biol. Med.*, vol. 111, no. 3, pp. 544–547, 1962, doi: 10.3181/00379727-111-27849.
- [29] S. M. Ali and G. Yosipovitch, "Skin pH: From basic science to basic skin care," *Acta Derm. Venereol.*, vol. 93, no. 3, pp. 261–267, 2013, doi: 10.2340/00015555-1531.
- [30] I. Giannopoulou, F. Saïs, and R. Thomopoulos, "Linked data annotation and fusion driven by data quality evaluation," *Rev. des Nouv. Technol. l'Information*, vol. E.28, pp. 257–262, 2015.
- [31] C. W. Li *et al.*, "Anti-inflammatory property of the ethanol extract of the root and rhizome of Pogostemon cablin (Blanco) benth," *Sci. World J.*, vol. 2013, 2013, doi: 10.1155/2013/434151.
- [32] J. C. Fehrenbacher, M. R. Vasko, and D. B. Duarte, "Models of inflammation: carrageenan-or complete freund's adjuvant (CFA)-induced edema and hypersensitivity in the rat," *Curr. Protoc. Pharmacol.*, vol. 56, no. 1, pp. 4–5, 2012.
- [33] J. Y. Lee and W. Park, "Anti-inflammatory effect of myristicin on RAW 264.7 macrophages stimulated with polyinosinic-polycytidylic acid," *Molecules*, vol. 16, no. 8, pp. 7132–7142, 2011, doi: 10.3390/molecules16087132.
- [34] K. Dewi, B. Widyarto, P. Erawijantari, and W. Widowati, "In vitro study of Myristica fragrans seed (Nutmeg) ethanolic extract and quercetin compound as anti-inflammatory agent," *Int. J. Res. Med. Sci.*, vol. 3, no. 9, pp. 2303–2310, 2015, doi: 10.18203/2320-6012.ijrms20150621.
- [35] M. E. Lane, "Skin penetration enhancers," *Int. J. Pharm.*, vol. 447, no. 1–2, pp. 12–21, 2013, doi: 10.1016/j.ijpharm.2013.02.040.
- [36] R. M. Watkinson, C. Herkenne, R. H. Guy, J. Hadgraft, G. Oliveira, and M. E. Lane, "Influence of Ethanol on the Solubility, Ionization and Permeation Characteristics of Ibuprofen in Silicone and Human Skin," *Skin Pharmacol. Physiol.*, vol. 22, no. 1, pp. 15–21, 2009, doi: 10.1159/000183922.

April 20th, 2021

Dear Editor-in-Chief

International Journal of Public Health Science (IJPHS)

We would like to appreciate the time and effort that the reviewers dedicated to providing feedback on our manuscript and are grateful for the insightful comments on and valuable improvements to our paper. We have attached a revised manuscript of Anti-Inflammatory Activity of Indonesia nutmeg seeds: A Topical Gel Formulation with Reference ID Number: 20921.

No conflict of interest exits in the submission of this manuscript, and manuscript is approved by all authors for publication. We would like to declare on behalf of my coauthors that the work described was original research which has not been published previously, and not under considerations for publication elsewhere, in whole or in part. All the authors listed have approved the manuscript that is enclosed.

In this work, we have revised our final manuscript according to the reviewer's suggestions. The change is highlighted in green color within the manuscript. We hope this revision is suitable for "International Journal of Public Health Science".

We deeply appreciate your consideration of our manuscript. If you have any queries, please don't hesitate to contact us at the address below.

Yours Sincerely,

Corresponding author:

apt. Azis Ikhsanudin, M.Sc.,

Assistant Professor, Department of Pharmaceutics

Ahmad Dahlan University

Prof Dr Soepomo Warungboto Umbulharjo, Yogyakarta, Indonesia

Email: azis.ikhsanudin@pharm.uad.ac.id

Tel: +62 89506685862

## Response to reviewers

#### **Reviewer #1 comment:**

TITLE

## 1. Reviewer's comment:

The title cannot be longer than 10 words!

## **Author's response:**

Thank you for pointing this out. We have revised the research title with no longer than 10 words. The revised title is:

"Anti-Inflammatory Activity of Indonesia nutmeg seeds: A Topical Gel Formulation"

## **ABSTRACT**

## 1. Reviewer's comment:

Abstract cannot be longer than 200 words

## **Author's response:**

We have revised our research abstract with less than 200 words (198 words)

## **ABSTRACT**

Herbal medicines have been shown as anti-inflammatory with potentially lesser side effects. The active compound of nutmeg seed is proven to accelerate the healing process of inflammation. This study aims to evaluate the gel formulation of Indonesian nutmeg seed extract for anti-inflammatory activity. A true experimental posttest only with control group design was used in this study. The gel was formulated with various concentrations of nutmeg seed extract, namely formulations F1 (0%), F2 (2%), F3 (4%), F4 (8 %), and F5 (12%). Analysis of variance (ANOVA) followed by the least significant difference (LSD) methods were performed with SPSS version 22. The results showed that all formulas had an opaque physical appearance, brownish-yellow color, soft texture, and aromatic odor. The increase of extract concentration in gel formula will affect the adhesion and spreadability. F5 showed the highest anti-inflammatory activity compared to other groups. This formula was generally identified as having a good physical appearance, homogeneity, and stability with a pH value of  $6.16 \pm 0.24$ , adhesiveness of  $51.12 \pm 0.15$  sec, and a spreadability of  $19.54 \pm 0.12$  cm2. Therefore, Indonesian nutmeg has the potential to be well-acceptable as a candidate for topical anti-inflammatory agents in global health benefits

## 2. Reviewer's comment:

What research design does this research use?

## **Author's response:**

We have added our research design in the abstract. The revised sentence have been marked in yellow block color.

Herbal medicines have been shown as anti-inflammatory with potentially lesser side effects. The active compound of nutmeg seed is proven to accelerate the healing process of inflammation. This study aims to evaluate the gel formulation of Indonesian nutmeg seed extract for anti-inflammatory activity. A true experimental posttest only with control group design was used in this study. The gel was formulated with various concentrations of nutmeg

seed extract, namely formulations F1 (0%), F2 (2%), F3 (4%), F4 (8%), and F5 (12%). Analysis of variance (ANOVA) followed by the least significant difference (LSD) methods were performed with SPSS version 22. The results showed that all formulas had an opaque physical appearance, brownish-yellow color, soft texture, and aromatic odor. The increase of extract concentration in gel formula will affect the adhesion and spreadability. F5 showed the highest anti-inflammatory activity compared to other groups. This formula was generally identified as having a good physical appearance, homogeneity, and stability with a pH value of  $6.16 \pm 0.24$ , adhesiveness of  $51.12 \pm 0.15$  sec, and a spreadability of  $19.54 \pm 0.12$  cm2. Therefore, Indonesian nutmeg has the potential to be well-acceptable as a candidate for topical anti-inflammatory agents in global health benefits.

## RESEARCH METHOD

## 1. Reviewer's comment:

What research design does this research use?

## **Author's response:**

Thank you for the comment. We've also added the description of research design in the research method section.

## RESEARCH METHOD

The research method used true experimental post-test only with control group. This study aimed to compare the anti-inflammatory activity between formula group of topical herbal gel from Indonesian nutmeg seed extract.

#### CONCLUSION

## 1. Reviewer's comment:

implications for future research?

## **Author's response:**

Thanks for the comments regarding the implications in future research.

We have added in conclusion that further studies should implement a novel pharmaceutical technology in topical drug delivery system regarding to enhance the anti-inflammation activity. The revised sentences have been marked in yellow block color.

## **CONCLUSION**

These present study indicates that nutmeg gel extract (*Myristica fragrans*, Houtt) with 12% ethanolic extract concentration has a good physical appearance and potent anti-inflammatory activity than other formulas and marketed product. Further study should implement a novel pharmaceutical technology in topical drug delivery system regarding to enhance the anti-inflammation activity. Hence, Indonesian nutmeg seed extract could be a potential candidate as anti-inflammatory in topical gel dosage form application.

## **International Journal of Public Health Science (IJPHS)**

Vol. x, No. x, March 2020, pp. xx~xx

# Topical Anti-Inflammatory Activity of Herbal Gel Formulation Indonesian Nutmeg seeds (*Myristica fragrans*, Houtt.)

Azis Ikhsanudin<sup>1\*</sup>, Lolita Lolita<sup>2</sup>, Daniar Dyanas Rais<sup>3</sup>

<sup>1,2,3</sup>Faculty of Pharmacy, Universitas Ahmad Dahlan, Yogyakarta Province 55164, Indonesia <sup>3</sup>Research Division of Clinical Pharmacology, the First Affiliated Hospital of Nanjing Medical University, P.R. China

#### **Article Info**

#### Article history:

Received Revised Accepted

## Keywords:

Nutmeg seed Myristicin Herbal gel Anti-inflammatory

## **ABSTRACT**

Cutaneous use of synthetic non-steroidal anti-inflammatory drugs may result in mucosal skin damage. To date, herbal medicines have been shown as anti-inflammatory with potentially lesser side effects. The active compound of nutmeg seed is proven to accelerate the healing process of inflammation. This study aims to determine and evaluate the gel formulation of Indonesian nutmeg seed extract for anti-inflammatory activity. The gel was formulated with various concentrations of nutmeg seed extract, namely formulations F1 (0%), F2 (2%), F3 (4%), F4 (8 %), and F5 (12%). Analysis of variance (ANOVA) followed by the least significant difference (LSD) methods were performed with SPSS version 22 to compare statistically significant differences between formula groups. The results showed that all formulas had an opaque physical appearance, brownish-yellow color, soft texture, and aromatic odor. The increase in extract concentration in gel formula will affect the adhesion and spreadability. Formulation F5 showed the highest anti-inflammatory activity compared to other groups. This formula was generally identified as having a good physical appearance, homogeneity, and stability with a pH value of 6.16  $\pm$  0.24, adhesiveness of 51.12  $\pm$ 0.15 sec, and a spreadability of  $19.54 \pm 0.12$  cm<sup>2</sup>. Therefore, Indonesian nutmeg has the potential to be well-acceptable as a candidate for topical anti-inflammatory agents in global health benefits

This is an open access article under the <u>CC BY-SA</u> license.



## Corresponding Author:

apt. Azis Ikhsanudin, M.Sc Faculty of Pharmacy, Ahmad Dahlan University,

Prof Dr Soepomo Warungboto Umbulharjo, Yogyakarta, Indonesia

Email: azis.ikhsanudin@pharm.uad.ac.id

## 5. INTRODUCTION.

Inflammation is a tissue-response cascade defense mechanism for specific inflammatory stimuli or pathogenic infections. This inflammatory reaction involves different cellular and vascular pathways with specific humoral secretions to prevent damage and restore tissue function [1]. Specific receptor cells could also trigger various pro and anti-inflammatory cytokines such as IL-1b, IL-6, IL-1a, and TNF- $\alpha$ , which modulate neutrophil migration to endothelial cells. Neutrophils will be released into circulation upon encountering multiple danger signals by producing inflammatory cytokines. Moreover, monocytes will differentiate into tissue macrophages by phagocytosis of damaged cells and other invading pathogens [2][3][4].

Journal homepage: http://ijphs.iaescore.com

The drugs most commonly used for alleviating acute and chronic inflammation are non-steroidal anti-inflammatory drugs (NSAIDs) and steroids (corticosteroids)[5][6]. These medications are typically used in combination because of their slightly distinct actions in granulation tissue. NSAIDs inhibit the cyclooxygenase enzymes (COX-1 and COX-2) responsible for regulating many cellular processes during the inflammatory response [7]. Steroids could suppress inflammation intensely by binding to receptors that control phospholipase A2, COX-2, iNOS, and interleukin. Unfortunately, two classes of drugs should be taken with caution because of the potential adverse events, e.g., gastrointestinal bleeding, myocardial infarction, and kidney disorders [8][9][10].

The development of a plant-based drug is currently the primary alternative approach regarding the long-term risk of using anti-inflammatory drugs. Under these circumstances, medicinal plants have significantly rediscovered cellular pathways to provide potent active compounds as therapeutic phytochemicals [11]. Furthermore, the market for herbal medicines accounts for 83% worldwide in the treatment of inflammatory diseases. It is estimated that it will reach a value of approximately more than 95% in the forthcoming years due to increased revenues from these preparations. Indonesia is the largest producer and exporter of nutmeg in the global market. Overall export volume tended to increase by an average of 3.07% per year during 2009-2018 period [12]. Indonesia nutmeg is of good quality due to its low and almost non-existent aflatoxin content [13]. The myristin content in Indonesian nutmeg was 8.72% higher than Indian nutmeg, which was only 3.8% [14], [15].

A feasibility study reports that myristicin has been recognized in pharmacological mechanisms as a potent anti-inflammatory activity. Several studies have shown that nutmeg seed essential oil containing myristicin and aluminometasilicate was useful for anti-inflammatory activity. It works by inhibiting the biosynthesis of prostaglandin in the human colon. The high myristicin content in nutmeg could play a role in reducing TNF- $\alpha$  levels. Myristicin also demonstrates a similar mechanism as NSAIDs such as indomethacin and aspirin [16][17][18][19].

Topical drug delivery systems are gaining popularity in developing local and systemic drug delivery systems. This system could avoid gastrointestinal irritation, overcome the "first pass" effect and maximize drug concentrations at receptor site [20]. Furthermore, the gel has biodegradable, biocompatible, consistent properties, reasonable penetration rates, and longer retention time [21][22][23]. Previous studies have reported the topical gel formulation of ethanol extract of nutmeg using the albumin denaturation method. This finding showed that nutmeg extract gel could inhibit albumin's denaturation higher than the marketed synthetic drug gels [24]. In our recent study, we used the carrageenan-induced rat paw edema model to evaluate anti-inflammatory activity. The measurement of anti-inflammatory activity was based on the increase in the volume of rat leg edema, and the percentage was calculated based on the AUC value. This approach is more representative for determining the actual anti-inflammatory activity for topical drug preparations. Therefore, our study aimed to design and evaluate Indonesian nutmeg seed extract's topical anti-inflammatory activity using the carrageenan-induced rat paw edema model. This study's significance is expected to produce potent plant-based anti-inflammatory properties based on good efficacy and acceptance standards.

## 6. RESEARCH METHOD

## 2.3 Plant Materials and Chemical Reagents

We collected fresh nutmeg seeds from Malikrubu Regency, Ternate, North Maluku, Indonesia. The plant was identified and approved by the Biology Laboratory of the Faculty of Mathematics and Natural Sciences, Ahmad Dahlan University. The chemical reagents such as distilled water, carbopol 940, methylparaben, propylene glycol, and triethanolamine were obtained through Brataco, Ltd, Indonesia. Voltaren gel containing 1% diclofenac sodium was obtained from the community pharmacist store, Ahmad Dahlan University. All of the reagents were of analytical and pharmaceutical grade.

## 2.4 The Extraction of Indonesian Nutmeg Seed

The purpose of the extraction was to preserve myristicin as an anti-inflammatory bioactive compound in nutmeg seed. Sample preparation was accomplished by peeling the fresh nutmeg using sharp stainlesssteel knives. Afterward, nutmeg seeds should be washed and dried in an oven at  $40^{\circ}$ C for 48 hours. The dried nutmeg seed should be ground to 0.4mm in size using an electric blender to enhance the contact surface. The seed extract was prepared by maceration using 70% ethanol as a solvent for three days at room temperature. About 500 g of nutmeg seed powdered was loaded using 1 liter of 70% ethanol for maceration, followed by stirring with a magnetic stirrer for 2 hours. The supernatant liquid mixture was poured onto filter paper and allowed to stand for 24 hours before being filtered. The whole liquid was filtered using a Buchner porcelain funnel. This treatment procedure was repeated for 3 x 24 hours to obtain a clear filtrate. Further, a gradual evaporation process was performed to obtain a concentrated extract [25].

## 2.3 Topical Gel Formulation of Indonesian Nutmeg Seed Extract

The gels were formulated using different concentration of nutmeg seeds extract ethanol (0%; 2%; 4%; 8% and 12%), carbopol 940 (1.5%), ethanol 96% (4%), propylene glycol (10%), glycerin (20%), triethanolamine (0.8%), methylparaben (0.1%), polysorbate 80 (0.8%), and purified water (q.s to 100%). We prepared the gelbased mixture by mixing carbopol in purified water and maintained it with magnetic stirring until homogeneous (mixture A). Propylene glycol and triethanolamine are dissolved in mixture A to obtain a swollen gel with a pH of 5-6 (mixture B). Afterward, add the nutmeg ethanol extract to mixture B and stir with methylparaben to form a stable and homogeneous gel. Furthermore, the nutmeg extract gels must be stored in a tightly-closed container for 24 hours until the bubbles are removed [24]. The ethanol extract gel formula of nutmeg seed was shown in Table 1

Table 1. Gel formula of ethanol extract nutmeg seed

Ingredient			Formula (gram)		
	F1	F2	F3	F4	F5
Nutmeg seed extract	0	1	2	4	6
Carbopol 940	0.75	0.75	0.75	0.75	0.75
Ethanol 96%	2	2	2	2	2
Propylenglycol	5	5	5	5	5
Glyserin	10	10	10	10	10
Trietanolamin	0.4	0.4	0.4	0.4	0.4
Methyl paraben	0.05	0.05	0.05	0.05	0.05
Polysorbate 80	0.4	0.4	0.4	0.4	0.4
Purified water ad	50	50	50	50	50

Note: The positive control (F6) used voltaren 50 g gel containing 1% diclofenac sodium

## 2.4 Evaluation of Formulated Nutmeg Seed Gel

The formulated nutmeg seed gel was characterized by specific physical properties such as organoleptic, pH, spreadability, and adhesivity. This parameter has the function of assessing the quality of the gel in various formulas. Another parameter was to investigate the potential activity of the gel as an anti-inflammatory. The test procedure was performed using the carrageenan-induced rat paw edema model. This acute inflammation model is well-accepted and has long been used to determine its potent anti-inflammatory effect.

## 2.4.1. Organoleptic properties

Gel formulations with or without nutmeg seed extract were evaluated organoleptically for color, texture, and homogeneity. These characteristics were performed by visual observation. The formulation consistency and the particle's coarseness were checked by texture and homogeneity parameters. To assess these parameters, we pressed a small amount of formulated gel between the thumb and the index finger.

## 2.4.2 pH Measurement

Electrochemical pH measurement is used to determine the acidity or alkalinity through a digital pH meter (HANNA HI9813-6 Portable). The principle step was to weigh 1 gram of each gel formula and completely immersed it in the glass electrode. All measurements were taken three times, for which the average reading was recorded [26].

## 2.4.3 Spreadability

The prepared gel spreadability was intended to evaluate the gel's ability to spread and absorb on the skin. This technique was performed by measuring the diameter of spread of 1 gram of gel samples on two horizontal glass slides (10x20cm2) after 60 sec. The standard upper plate for determining gel spreadability was 0.5 g. Each gel formulation was recorded in triplicate [24].

## 2.4.4 Adhesivity

The purpose of gel adhesion testing was to determine the strength that separates the gel from the surface. A tensile adhesion test measures this by placing 2 grams of gel between two resin plates in the designated area. Then clamp the load of 1 kg on the glass plate for 5 minutes. The static resin plate was attached to the adhesion apparatus by applying a trigger load of 80 grams. Further, the evaluation was carried out to determine the length of time using a stopwatch until the resin plate runs out [27].

## 2.4.5 In-vitro anti-inflammatory activity of nutmeg seed extract

The study protocol has obtained ethical approval from Ahmad Dahlan University Ethics Committee with reference number 011502014. The anti-inflammatory properties of nutmeg seed gel products were measured using carrageenan-induce acute inflammation model in hind leg edema of male Wistar rat. The carrageenan-induced paw edema method is a well-defined model of acute inflammation involving various inflammatory mediators, as described by Winter et al. [28]. Initially, healthy Wistar strains (150-200 grams per each) were obtained from an animal breeding house, Faculty of Veterinary Medicine, Gadjah Mada University, Yogyakarta. Rats kept under laboratory conditions were fed twice daily with standard rat chow and water ad libitum. All rats were randomly allocated into six groups: one negative control group, four intervention groups,

and one positive control group. Each group was comprising at least five experimental animals. Before the experiment, the animal had to fast for 24 hours with access to water. 50  $\mu$ l of 1%  $\lambda$ -carrageenan solution (Sigma-Aldrich, Milan, Italy) was suspended in 1% NaCl solution, which had been prepared 1 hour before the experiment. Furthermore, this solution should be injected into the plantar side of the rat's right hind paw. About 0.2 grams of nutmeg seed gel in a different formula was gently applied 50 times with the index finger after 15 minutes of carrageenan injection. The rat paw edema volume was measured immediately using a plethysmometer (model UGO BASILE S.R.L: 7141) at 15 minutes to 3 hours after carrageenan injection. The percentage of inflammation was calculated by the following formula:

% Inflammatory = 
$$\frac{\text{V0 - Vt}}{\text{V0}} \times 100\%$$

**Note**: Vo and Vt are the volumes of rat paw edema from the control group and experimental group.

## 2.5 Data Analysis

The study results were reported descriptively on several test data such as organoleptic properties, pH, adhesiveness, and spreadability. The statistical analysis was performed using One-Way ANOVA and followed by the LSD-post hoc test. A *p*-value less than 0.05 was considered statistically significant.

## 7. RESULTS AND DISCUSSIONS

We have successfully formulated herbal gel from the ethanol extract of nutmeg seed in various formulas. Our study demonstrated that the obtained nutmeg seed gel has a good physical appearance and an anti-inflammatory activity due to acute edema reduction. The detailed finding regarding physical characteristics and anti-inflammatory activity will be presented in the subsection below.

## 3.1 Organoleptic properties

As shown in Table 2, the nutmeg seed gel has a good cosmetic appearance with a soft texture, opaque, brownish-yellow in color, characteristic aromatic odor, and homogeneous without segregation. In general, the dense pigment of nutmeg seed contains more dyes to produce a brownish-yellow gel. Previous studies have also reported that the nutmeg seed gel's physical properties were brownish yellow, fragrant and homogeneous[27][24]. The ethanolic extract does not dissolve completely in the gel bases due to the differences in solubility level.

Table 2. Organoleptic properties of nutmeg seed extract gels

Formula	Physical	Color	Odor	Texture	Homogenity	Segregation
	appearance					
1	Transparant	Clear white	Typical of base	Soft	Homogenous	No
2	Opaque	Brownish yellow	Typical of nutmeg oil	Soft	Homogenous	No
3	Opaque	Brownish yellow	Typical of nutmeg oil	Soft	Homogenous	No
4	Opaque	Brownish yellow	Typical of nutmeg oil	Soft	Homogenous	No
5	Opaque	Brownish yellow	Typical of nutmeg oil	Soft	Homogenous	No

The appearance of nutmeg seed extract gel with various extract concentrations is shown in Figure 1.



Figure 1. The physical appearance of nutmeg seed extract gel at F2 (2%); F3 (4%); F4 (8%) and F5 (12%) (from left to right)

## 3.2 pH Measurement

Each nutmeg seed extract gel formula shows the following pH value F1(4.69  $\pm$  0.74), F2 (5.38  $\pm$  0.78), F3 (5.82  $\pm$  0.15), F4 (6.33  $\pm$  0.13), F5 (6.16  $\pm$  0.24) and F6 (5.87  $\pm$  0.12) (Table 3). These pH value has met the standard criteria for topical skin administration (pH ranged 4-7). pH value measurement could be related to the safety and efficacy of topical drug preparations. Gel acidity and alkalinity will affect skin irritation, penetration and stability of active compounds into the skin. The more acidic the pH value of gel will increase skin irritation, while the more alkaline it will cause dry skin [29][27]. The statistic analysis using the LSD post hoc test showed a significant difference in pH value between F3 to F5 compared to F1 (p<0.001). These results mean that the pH value will increase with the addition of nutmeg extract concentration in the gel base. Meanwhile, the pH value of F1 (gel base) was not significantly different from F2 (extract 2%) (p= 0.121). This defines that both formulas have a similar pH value. As a gelling agent, each end of carbopol's chain has an

acidic carboxylic chemical structure when reacting to water. Carbopol 940 is easily ionized during the neutralization process with the addition of triethanolamine. Therefore, the interaction between carbopol 940 and triethanolamine could affect the pH stability based on the increase in extract concentration [30].

Table 3. The pH value of nutmeg seed extract gel

Formula	pH (Mean $\pm$ SD)		p-v	ralue (LSD post l	noc test)	
		F1	F2	F3	F4	F5
F1	4.69±0.74	-	0.121	0.020*	0.002*	0.050*
F2	5.38±0.78	0.121	-	0.306	0.041*	0.084
F3	5.82±0.15	0.020*	0.306	-	0.234	0.419
F4	6.33±0.13	0.002*	0.041*	0.236	-	0.680
F5	6.16±0.24	0.005*	0.084	0.419	0.680	-

Note: \* p-value less than 0.05

## 3.3 Adhesivity

Penetration of the active compound gel into the skin will increase along with the enhancement of gel adhesivity properties. The results showed that the average adhesion score of nutmeg seed extract gel was F1 (13.57 $\pm$ 0.11), F2 (18.32 $\pm$ 0.29), F3 (44.33 $\pm$ 0.09), F4 (56.54 $\pm$ 0.19), and F5 (51.12 $\pm$ 0.15) (Table 4). Based on a one-way ANOVA test followed by LSD statistical analysis, the nutmeg extract gel of F4 was significantly greater in adhesion than other formulas (p<0.005). Furthermore, the formulation of F5 with 12% nutmeg seed ethanol extract showed a decrease in adhesiveness compared to F4. This decrease might be due to the pH value being smaller at F5 than F4. The cross-linking between carbopol and other molecules, such as solvents, could also result in ionic attraction, and increasing the gel's viscosity. In acid conditions, increasing the gel's viscosity will have an impact on decreasing the adhesive properties.

Table 4. Adhesive properties of the nutmeg seed extract gel

	Mean $\pm$ SD (second)					
Parameter	F1	F2	F3	F4	F5	<b>1</b>
Adhesivity test (sec)	13.57±0.11	18.32±0.29	44.33±0.09	56.54±0.19	51.12±0.15	0.000

## 3.4 Spreadability

As shown in Table 5, the average spreadability of the nutmeg gel formulations showed F1 (9.38 $\pm$ 0.65), F2 (11.54 $\pm$ 0.21), F3 (13.39 $\pm$ 0.06), F4 (14.85 $\pm$ 0.34), and F5 (19.54 $\pm$ 0.12). These findings concluded that increasing the gel extract concentration would affect the broadest spreadability of the gel. The formula F5 (12%) has the greatest spreadability compared to other formulas. The statistical analysis results through one-way ANOVA stated a significant difference in spreadability between all formulas (p<0.005). Carbopol 940, an acrylic polymer gelling agent, could regulate the viscosity in 1-2% concentrations by producing a three-dimensional matrix to form a viscous gell. The spreadability of the gel formula was highly dependent on the viscosity of the carbopol. The less viscous gelling agent will enhance the spreadability of the gel. The swelling gel process occurs due to the solvent penetration, leading to the cross-linked polymer network to maintain the dosage form and the binding drug particles. However, the alkaline environment pH will increase the density of negative charge, resulting in the gel swollen and facilitating the drug's release [21][30]. Furthermore, the addition of nutmeg seed extract to the gel base will also affect the formation of hydrogen bonds in the physical cross-linking, which influences a reduced viscosity [21].

Table 5. Spreadability properties of the nutmeg seed extract gel

		Mean $\pm$ SD (cm <sup>2</sup> )					
Parameter	F1	F2	F3	F4	F5	<i>p</i> -value	
Spreadability test (cm)	9.38±0.65	11.54±0.21	13.39±0.06	14.85±0.34	19.54±0.12	0.000	

## 3.5 In-vitro anti-inflammatory activity of nutmeg seed extract gel

The evaluation of the nutmeg seed herbal gel's anti-inflammatory activity was observed within 180 min (Figure 2). During this time, carrageenan will significantly increase TNF-α, IL-1β, PGE2, iNOS, and COX-2 proteins in peripheral leg inflammation. Carrageenan-induced leg edema follows a model of acute inflammation which consists of two phases: first, which was detected after about 1 hour and was called the fast phase, with the release of histamine and serotonin, and the second stage was called the late phase with the mediators (kinins, prostaglandins) released after 2 and 3 hours, respectively [31]. A previous study suggested that intraplantar carrageenan injection will stimulate rat paw edema and release inflammatory mediators within 180 min [32]. This study is in line with our findings that decreased edema volume also occurred at 180 minutes after intraplantar carrageenan administration.

Figure 2 showed that the formulation of nutmeg seed extract gel has anti-inflammatory effects. It can be proven that the percentage volume of edema in each formula (F2 to F6) was lower than F1 (gel-based negative control). The anti-inflammatory activity of F5 (12%) was higher than that of F2 (2%) and F3 (4%). Furthermore, F4 (8%) and F5 (12%) had anti-inflammatory activity similar to F6 (positive control). This anti-inflammatory activity is influenced by the main compound contained in the nutmeg, namely myristicin. The active compound of nutmeg seed extract, myristicin, has a vital function as an anti-inflammatory by inhibiting chemokines, cytokines, nitrous oxide, and double-stranded growth factors RNA (dsRNA), which are stimulated by macrophages through calcium [33]. Other quercetin compounds also have anti-inflammatory activity by inhibiting the secretion of TNF- $\alpha$ , IL-6, IL-1 $\beta$ , and nitric oxide (NO) [34].

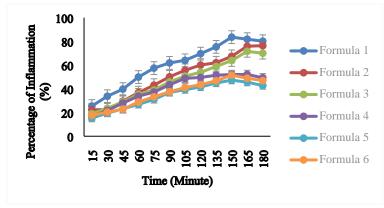


Figure 2. Inflammatory activity of nutmeg seed extract gels.

The volume increase in rat leg edema was used to calculate each formula's anti-inflammatory percentage based on the AUC value. The calculated AUC value and the percentage of anti-inflammatory can be seen in Table 6. Based on the LSD post hoc test analysis reported that the anti-inflammatory activity of F4, F5 did not have a significant difference compared to F6 (positive control). It was concluded that F4 and F5 had similar anti-inflammatory activity as the positive control (1% diclofenac sodium gel in the marketed product). Furthermore, the anti-inflammatory activity of F5 (44.92%) was higher significant difference compared to F4 (37.87%) with p=0.025 The AUC calculation results showed that F5 had the lowest AUC value (5803.72) with the highest percentage of anti-inflammatory activity (44.92%) compared to other formulas (F1 to F5).

Table 6. The percentage of anti-inflammatory activity of nutmeg extract gel was compared between groups.

Formula	AUC Anti-inflammatory activity (%)	p-value (LSD post hoc test)						
		activity (%)	F1	F2	F3	F4	F5	F6
F1	10536.94	0	-	0.000*	0.000*	0.000*	0.000*	0.000*
F2	9304.97	11.69	0.000*	-	0.000*	0.000*	0.000*	0.000*
F3	7919.26	24.84	0.000*	0.000*	-	0.000*	0.000*	0.000*
F4	6546.62	37.87	0.000*	0.000*	0.000*	-	0.025*	0.423
F5	5803.727	44.92	0.000*	0.000*	0.000*	0.025*	-	0.124
F6	6295.140	40.25	0.000*	0.000*	0.000*	0.423	0.124	-

Note: \* p-value less than 0.05; F6 is positive control

The highest anti-inflammatory activity of F5 was due to the gel's spreadability and its adhesion to the skin surface. F5 has a good adhesiveness of  $(51.12 \pm 0.15)$  sec with a spreadability of  $(19.54 \pm 0.12)$  cm. The topical drug penetration into the skin was affected by the length of time the gel is in contact with the skin surface. F5 has the highest concentration of nutmeg seed extract (12%) than other formulas (F2 to F4). The high content of myristicin in the F5 gel formula will improve its anti-inflammatory effect by enhancing the drug permeation into the skin. Besides, F5 also had the optimal ternary system composition between ethanol, propylene glycol, and purified water. Propylene glycol (PG) as an enhancer will affect the drug absorption into the skin [35]. A study of topical ibuprofen on human skin reported that ethanol's ternary solvent system, propylene glycol, and water had higher permeability into the skin than the binary solvent system. The ternary system will increase the solubility partition and membrane permeability partitioning by maximizing the active drug flux [36].

## 8. CONCLUSION

These present study indicates that nutmeg gel extract (*Myristica fragrans*, Houtt) with 12% ethanolic extract concentration has a good physical appearance and potent anti-inflammatory activity than other formulas

and marketed product. Hence, nutmeg seed extract could be considered as a potential candidate for antiinflammatory in topical application.

## ACKNOWLEDGEMENTS

The author would like to thank the whole researcher in Ahmad Dahlan University who has supported this research.

## REFERENCES

- [1] S. Kany, J. T. Vollrath, and B. Relja, "Cytokines in inflammatory disease," *Int. J. Mol. Sci.*, vol. 20, no. 23, pp. 1–31, 2019, doi: 10.3390/ijms20236008.
- Y. Su, J. Gao, P. Kaur, and Z. Wang, "Neutrophils and macrophages as targets for development of nanotherapeutics in inflammatory diseases," *Pharmaceutics*, vol. 12, no. 12, pp. 1–24, 2020, doi: 10.3390/pharmaceutics12121222.
- [3] A. L. Loktionov, A. I. Konoplya, M. A. Lunev, and A. V. Karaulov, "Immune and oxidant disorders in the pathogenesis of inflammatory periodontal diseases," *Immunologiya*, vol. 36, no. 5, pp. 319–328, 2015, doi: 10.1111/prd.12002.Inflammatory.
- [4] A. Tedgui and Z. Mallat, "Anti-inflammatory mechanisms in the vascular wall," *Circ. Res.*, vol. 88, no. 9, pp. 877–887, 2001, doi: 10.1161/hh0901.090440.
- [5] M. Del Grossi Moura, L. C. Lopes, M. T. Silva, S. Barberato-Filho, R. H. L. Motta, and C. De Cássia Bergamaschi, "Use of steroid and nonsteroidal anti-inflammatories in the treatment of rheumatoid arthritis: Systematic review protocol," *Med. (United States)*, vol. 97, no. 41, 2018, doi: 10.1097/MD.000000000012658.
- [6] A. Lahoti, B. S. Kalra, and U. Tekur, "Evaluation of the analgesic and anti-inflammatory activity of fixed dose combination: non-steroidal anti-inflammatory drugs in experimental animals.," *Indian J. Dent. Res.*, vol. 25, no. 5, pp. 551–554, 2014, doi: 10.4103/0970-9290.147071.
- [7] K. D. Brandt, "The mechanism of action of nonsteroidal antiinflammatory drugs," *J. Rheumatol.*, vol. 18, no. SUPPL. 27, pp. 120–121, 1991, doi: 10.5772/68090.
- [8] P. Kuropakornpong, A. Itharat, S. Panthong, S. Sireeratawong, and B. Ooraikul, "In Vitro and in Vivo Anti-Inflammatory Activities of Benjakul: A Potential Medicinal Product from Thai Traditional Medicine," *Evidence-based Complement. Altern. Med.*, vol. 2020, 2020, doi: 10.1155/2020/9760948.
- [9] K. R. Patil *et al.*, "Animal models of inflammation for screening of anti-inflammatory drugs: Implications for the discovery and development of phytopharmaceuticals," *Int. J. Mol. Sci.*, vol. 20, no. 18, 2019, doi: 10.3390/ijms20184367.
- [10] S. Tsurufuji, K. Sugio, H. Sato, and K. Ohuchi, "A review of mechanism of action of steroid and non-steroid anti-inflammatory drugs," *Inflamm. Mech. Treat.*, no. 202, pp. 63–78, 1980, doi: 10.1007/978-94-010-9423-8 7.
- [11] V. P. Gullo, J. McAlpine, K. S. Lam, D. Baker, and F. Petersen, "Drug discovery from natural products," *J. Ind. Microbiol. Biotechnol.*, vol. 33, no. 7, pp. 523–531, 2006, doi: 10.1007/s10295-006-0107-2.
- [12] H. Juliani Purba, E. Supriadi Yusufi, and J. Hestina, "Performane and Competitiveness of Indonesian Nutmeg in Export Market," *E3S Web Conf.*, vol. 232, p. 02018, 2021, doi: 10.1051/e3sconf/202123202018.
- [13] J. Marmis, N. S. Tongkeles, Lady Lengkey, and S. Ratulangi, "the Quality of Nutmeg Seeds and Mace," vol. 2, pp. 111–121, 2015.
- [14] K. Karmanah, S. Susanto, W. D. Widodo, and E. Santosa, "The Fruit Characteristics of Ambon Forest Nutmeg (Myristica fatua Houtt) and Banda Nutmeg (Myristica fragrans Houtt)," *J. Ilmu Pertan. Indones.*, vol. 25, no. 2, pp. 292–300, 2020, doi: 10.18343/jipi.25.2.292.
- [15] I. P. S. Kapoor, B. Singh, G. Singh, C. S. De Heluani, M. P. De Lampasona, and C. A. N. Catalan, "Chemical composition and antioxidant activity of essential oil and oleoresins of nutmeg (Myristica fragrans Houtt.) fruits," *Int. J. Food Prop.*, vol. 16, no. 5, pp. 1059–1070, 2013, doi: 10.1080/10942912.2011.576357.
- [16] I. Matulyte *et al.*, "The essential oil and hydrolats from myristica fragrans seeds with magnesium aluminometasilicate as excipient: Antioxidant, antibacterial, and anti-inflammatory activity," *Foods*, vol. 9, no. 1, 2020, doi: 10.3390/foods9010037.
- [17] Jinous Asgarpanah, "Phytochemistry and pharmacologic properties of Myristica fragrans Hoyutt.: A review," *African J. Biotechnol.*, vol. 11, no. 65, 2012, doi: 10.5897/ajb12.1043.
- [18] O. A. Olajide, J. M. Makinde, and S. O. Awe, "Evaluation of the pharmacological properties of nutmeg oil in rats and mice," *Pharm. Biol.*, vol. 38, no. 5, pp. 385–390, 2000, doi: 10.1076/phbi.38.5.385.5976.
- [19] W. K. Zhang et al., "Nutmeg oil alleviates chronic inflammatory pain through inhibition of COX-2

- expression and substance P release in vivo," *Food Nutr. Res.*, vol. 60, no. April, 2016, doi: 10.3402/fnr.v60.30849.
- [20] M. M. Ghareeb, "Design and in vitro characterization of a topical nanoemulsionenriched hydrogel of econazole nitrate," *J. Appl. Pharm. Sci.*, vol. 9, no. 1, pp. 51–57, 2019, doi: 10.7324/JAPS.2019.90108.
- [21] S. A. A. Nabi, M. A. Sheraz, S. Ahmed, N. Mustaan, and I. Ahmad, "Pharmaceutical Gels : A Review," *RADS J. Pharm. Pharm. Sci.*, vol. 4, no. 1, pp. 40–48, 2016.
- [22] C. Tipa, M. T. Cidade, T. Vieira, J. C. Silva, P. I. P. Soares, and J. P. Borges, "Article a new long-term composite drug delivery system based on thermo-responsive hydrogel and nanoclay," *Nanomaterials*, vol. 11, no. 1, pp. 1–22, 2021, doi: 10.3390/nano11010025.
- [23] R. Sánchez-Sánchez *et al.*, "Development of Hydrogel with Anti-Inflammatory Properties Permissive for the Growth of Human Adipose Mesenchymal Stem Cells," *J. Nanomater.*, vol. 2016, 2016, doi: 10.1155/2016/8654937.
- [24] S. Surendran and A. V. Vidyapeetham, "TOPICAL GEL AND ITS IN VITRO EVALUATION FOR ANTINFLAMMATORY Available Online through Research Article," no. April 2016, 2018.
- [25] E. F. Al-Jumaily and M. H. A. Al-Amiry, "Extraction and Purification of Terpenes from Nutmeg (myristica fragrans)," *J. Al-Nahrain Univ. Sci.*, vol. 15, no. 3, pp. 151–160, 2012, doi: 10.22401/jnus.15.3.21.
- [26] S. Karastogianni, S. Girousi, and S. Sotiropoulos, *pH: Principles and Measurement*, 1st ed., no. December. Elsevier Ltd., 2015.
- [27] R. Aiyalu, A. Govindarjan, and A. Ramasamy, "Formulation and evaluation of topical herbal gel for the treatment of arthritis in animal model," *Brazilian J. Pharm. Sci.*, vol. 52, no. 3, pp. 493–507, 2016, doi: 10.1590/s1984-82502016000300015.
- [28] C. A. Winter, E. A. Risley, and G. W. Nuss, "Carrageenin-Induced Edema in Hind Paw of the Rat as an Assay for Antiinflammatory Drugs," *Proc. Soc. Exp. Biol. Med.*, vol. 111, no. 3, pp. 544–547, 1962, doi: 10.3181/00379727-111-27849.
- [29] S. M. Ali and G. Yosipovitch, "Skin pH: From basic science to basic skin care," *Acta Derm. Venereol.*, vol. 93, no. 3, pp. 261–267, 2013, doi: 10.2340/00015555-1531.
- [30] I. Giannopoulou, F. Saïs, and R. Thomopoulos, "Linked data annotation and fusion driven by data quality evaluation," *Rev. des Nouv. Technol. l'Information*, vol. E.28, pp. 257–262, 2015.
- [31] C. W. Li *et al.*, "Anti-inflammatory property of the ethanol extract of the root and rhizome of Pogostemon cablin (Blanco) benth," *Sci. World J.*, vol. 2013, 2013, doi: 10.1155/2013/434151.
- [32] J. C. Fehrenbacher, M. R. Vasko, and D. B. Duarte, "Models of inflammation: carrageenan-or complete freund's adjuvant (CFA)-induced edema and hypersensitivity in the rat," *Curr. Protoc. Pharmacol.*, vol. 56, no. 1, pp. 4–5, 2012.
- [33] J. Y. Lee and W. Park, "Anti-inflammatory effect of myristicin on RAW 264.7 macrophages stimulated with polyinosinic-polycytidylic acid," *Molecules*, vol. 16, no. 8, pp. 7132–7142, 2011, doi: 10.3390/molecules16087132.
- [34] K. Dewi, B. Widyarto, P. Erawijantari, and W. Widowati, "In vitro study of Myristica fragrans seed (Nutmeg) ethanolic extract and quercetin compound as anti-inflammatory agent," *Int. J. Res. Med. Sci.*, vol. 3, no. 9, pp. 2303–2310, 2015, doi: 10.18203/2320-6012.ijrms20150621.
- [35] M. E. Lane, "Skin penetration enhancers," *Int. J. Pharm.*, vol. 447, no. 1–2, pp. 12–21, 2013, doi: 10.1016/j.ijpharm.2013.02.040.
- [36] R. M. Watkinson, C. Herkenne, R. H. Guy, J. Hadgraft, G. Oliveira, and M. E. Lane, "Influence of Ethanol on the Solubility, Ionization and Permeation Characteristics of Ibuprofen in Silicone and Human Skin," *Skin Pharmacol. Physiol.*, vol. 22, no. 1, pp. 15–21, 2009, doi: 10.1159/000183922.

## **BIOGRAPHIES OF AUTHORS (10 PT)**



Azis Iksanudin, is a lecturer in Faculty of Pharmacy, Ahmad Dahlan University, Yogyakarta Indonesia. His research interests are in the field of pharmaceutical technology, natural product, biomaterial, and drug delivery. He has many experience in conduct a researched granted from Higher Education of Indonesia and published some articles in Indonesian and scopus indexed International Journal.



Lolita is a Assistant Professor of Ahmad Dahlan University, Yogyakarta, Indonesia. She is also a Ph.D candidate from Nanjing Medical University, P.R. China. She has published some scientific articles related to social pharmacy, community health in accredited national journal of Indonesia. Her research interest in the topic of clinical pharmacy, pharmacogenomics, community health, and epidemiology. She is also a reviewer in scopus indexed international journals.



**Daniar Dyanas Rais** is an Indonesian pharmacist. She graduated from Bachelor degree programme in Faculty of Pharmacy, Ahmad Dahlan University. Now, she works as a pharmacist in Indonesian state-owned enterprise, Kimia Farma.

## **International Journal of Public Health Science (IJPHS)**

Vol. x, No. x, March 2020, pp. xx~xx

## Topical Anti-Inflammatory Activity of Herbal Gel Formulation Indonesian Nutmeg seeds (*Myristica fragrans*, Houtt.)

Azis Ikhsanudin<sup>1\*</sup>, Lolita Lolita<sup>2</sup>, Daniar Dyanas Rais<sup>3</sup>

<sup>1,2,3</sup>Faculty of Pharmacy, Universitas Ahmad Dahlan, Yogyakarta Province 55164, Indonesia <sup>3</sup>Research Division of Clinical Pharmacology, the First Affiliated Hospital of Nanjing Medical University, P.R. China

#### **Article Info**

#### Article history:

Received Revised Accepted

## Keywords:

Nutmeg seed Myristicin Herbal gel Anti-inflammatory

## **ABSTRACT**

Cutaneous use of synthetic non-steroidal anti-inflammatory drugs may result in mucosal skin damage. To date, herbal medicines have been shown as anti-inflammatory with potentially lesser side effects. The active compound of nutmeg seed is proven to accelerate the healing process of inflammation. This study aims to determine and evaluate the gel formulation of Indonesian nutmeg seed extract for anti-inflammatory activity. The gel was formulated with various concentrations of nutmeg seed extract, namely formulations F1 (0%), F2 (2%), F3 (4%), F4 (8 %), and F5 (12%). Analysis of variance (ANOVA) followed by the least significant difference (LSD) methods were performed with SPSS version 22 to compare statistically significant differences between formula groups. The results showed that all formulas had an opaque physical appearance, brownish-yellow color, soft texture, and aromatic odor. The increase in extract concentration in gel formula will affect the adhesion and spreadability. Formulation F5 showed the highest anti-inflammatory activity compared to other groups. This formula was generally identified as having a good physical appearance, homogeneity, and stability with a pH value of 6.16  $\pm$  0.24, adhesiveness of 51.12  $\pm$ 0.15 sec, and a spreadability of  $19.54 \pm 0.12$  cm<sup>2</sup>. Therefore, Indonesian nutmeg has the potential to be well-acceptable as a candidate for topical anti-inflammatory agents in global health benefits

This is an open access article under the CC BY-SA license.



## Corresponding Author:

apt. Azis Ikhsanudin, M.Sc Faculty of Pharmacy, Ahmad Dahlan University,

Prof Dr Soepomo Warungboto Umbulharjo, Yogyakarta, Indonesia

Email: azis.ikhsanudin@pharm.uad.ac.id

## 9. INTRODUCTION.

Inflammation is a tissue-response cascade defense mechanism for specific inflammatory stimuli or pathogenic infections. This inflammatory reaction involves different cellular and vascular pathways with specific humoral secretions to prevent damage and restore tissue function [1]. Specific receptor cells could also trigger various pro and anti-inflammatory cytokines such as IL-1b, IL-6, IL-1a, and TNF- $\alpha$ , which modulate neutrophil migration to endothelial cells. Neutrophils will be released into circulation upon encountering multiple danger signals by producing inflammatory cytokines. Moreover, monocytes will differentiate into tissue macrophages by phagocytosis of damaged cells and other invading pathogens [2][3][4].

Journal homepage: http://ijphs.iaescore.com

The drugs most commonly used for alleviating acute and chronic inflammation are non-steroidal anti-inflammatory drugs (NSAIDs) and steroids (corticosteroids)[5][6]. These medications are typically used in combination because of their slightly distinct actions in granulation tissue. NSAIDs inhibit the cyclooxygenase enzymes (COX-1 and COX-2) responsible for regulating many cellular processes during the inflammatory response [7]. Steroids could suppress inflammation intensely by binding to receptors that control phospholipase A2, COX-2, iNOS, and interleukin. Unfortunately, two classes of drugs should be taken with caution because of the potential adverse events, e.g., gastrointestinal bleeding, myocardial infarction, and kidney disorders [8][9][10].

The development of a plant-based drug is currently the primary alternative approach regarding the long-term risk of using anti-inflammatory drugs. Under these circumstances, medicinal plants have significantly rediscovered cellular pathways to provide potent active compounds as therapeutic phytochemicals [11]. Furthermore, the market for herbal medicines accounts for 83% worldwide in the treatment of inflammatory diseases. It is estimated that it will reach a value of approximately more than 95% in the forthcoming years due to increased revenues from these preparations. Indonesia is the largest producer and exporter of nutmeg in the global market. Overall export volume tended to increase by an average of 3.07% per year during 2009-2018 period [12]. Indonesia nutmeg is of good quality due to its low and almost non-existent aflatoxin content [13]. The myristin content in Indonesian nutmeg was 8.72% higher than Indian nutmeg, which was only 3.8% [14], [15].

A feasibility study reports that myristicin has been recognized in pharmacological mechanisms as a potent anti-inflammatory activity. Several studies have shown that nutmeg seed essential oil containing myristicin and aluminometasilicate was useful for anti-inflammatory activity. It works by inhibiting the biosynthesis of prostaglandin in the human colon. The high myristicin content in nutmeg could play a role in reducing TNF- $\alpha$  levels. Myristicin also demonstrates a similar mechanism as NSAIDs such as indomethacin and aspirin [16][17][18][19].

Topical drug delivery systems are gaining popularity in developing local and systemic drug delivery systems. This system could avoid gastrointestinal irritation, overcome the "first pass" effect and maximize drug concentrations at receptor site [20]. Furthermore, the gel has biodegradable, biocompatible, consistent properties, reasonable penetration rates, and longer retention time [21][22][23]. Previous studies have reported the topical gel formulation of ethanol extract of nutmeg using the albumin denaturation method. This finding showed that nutmeg extract gel could inhibit albumin's denaturation higher than the marketed synthetic drug gels [24]. In our recent study, we used the carrageenan-induced rat paw edema model to evaluate anti-inflammatory activity. The measurement of anti-inflammatory activity was based on the increase in the volume of rat leg edema, and the percentage was calculated based on the AUC value. This approach is more representative for determining the actual anti-inflammatory activity for topical drug preparations. Therefore, our study aimed to design and evaluate Indonesian nutmeg seed extract's topical anti-inflammatory activity using the carrageenan-induced rat paw edema model. This study's significance is expected to produce potent plant-based anti-inflammatory properties based on good efficacy and acceptance standards.

## 10. RESEARCH METHOD

## 2.5 Plant Materials and Chemical Reagents

We collected fresh nutmeg seeds from Malikrubu Regency, Ternate, North Maluku, Indonesia. The plant was identified and approved by the Biology Laboratory of the Faculty of Mathematics and Natural Sciences, Ahmad Dahlan University. The chemical reagents such as distilled water, carbopol 940, methylparaben, propylene glycol, and triethanolamine were obtained through Brataco, Ltd, Indonesia. Voltaren gel containing 1% diclofenac sodium was obtained from the community pharmacist store, Ahmad Dahlan University. All of the reagents were of analytical and pharmaceutical grade.

## 2.6 The Extraction of Indonesian Nutmeg Seed

The purpose of the extraction was to preserve myristicin as an anti-inflammatory bioactive compound in nutmeg seed. Sample preparation was accomplished by peeling the fresh nutmeg using sharp stainlesssteel knives. Afterward, nutmeg seeds should be washed and dried in an oven at  $40^{\circ}$ C for 48 hours. The dried nutmeg seed should be ground to 0.4mm in size using an electric blender to enhance the contact surface. The seed extract was prepared by maceration using 70% ethanol as a solvent for three days at room temperature. About 500 g of nutmeg seed powdered was loaded using 1 liter of 70% ethanol for maceration, followed by stirring with a magnetic stirrer for 2 hours. The supernatant liquid mixture was poured onto filter paper and allowed to stand for 24 hours before being filtered. The whole liquid was filtered using a Buchner porcelain funnel. This treatment procedure was repeated for 3 x 24 hours to obtain a clear filtrate. Further, a gradual evaporation process was performed to obtain a concentrated extract [25].

## 2.3 Topical Gel Formulation of Indonesian Nutmeg Seed Extract

The gels were formulated using different concentration of nutmeg seeds extract ethanol (0%; 2%; 4%; 8% and 12%), carbopol 940 (1.5%), ethanol 96% (4%), propylene glycol (10%), glycerin (20%), triethanolamine (0.8%), methylparaben (0.1%), polysorbate 80 (0.8%), and purified water (q.s to 100%). We prepared the gelbased mixture by mixing carbopol in purified water and maintained it with magnetic stirring until homogeneous (mixture A). Propylene glycol and triethanolamine are dissolved in mixture A to obtain a swollen gel with a pH of 5-6 (mixture B). Afterward, add the nutmeg ethanol extract to mixture B and stir with methylparaben to form a stable and homogeneous gel. Furthermore, the nutmeg extract gels must be stored in a tightly-closed container for 24 hours until the bubbles are removed [24]. The ethanol extract gel formula of nutmeg seed was shown in Table 1

Table 1. Gel formula of ethanol extract nutmeg seed

Ingredient			Formula (gram)		
	F1	F2	F3	F4	F5
Nutmeg seed extract	0	1	2	4	6
Carbopol 940	0.75	0.75	0.75	0.75	0.75
Ethanol 96%	2	2	2	2	2
Propylenglycol	5	5	5	5	5
Glyserin	10	10	10	10	10
Trietanolamin	0.4	0.4	0.4	0.4	0.4
Methyl paraben	0.05	0.05	0.05	0.05	0.05
Polysorbate 80	0.4	0.4	0.4	0.4	0.4
Purified water ad	50	50	50	50	50

Note: The positive control (F6) used voltaren 50 g gel containing 1% diclofenac sodium

## 2.4 Evaluation of Formulated Nutmeg Seed Gel

The formulated nutmeg seed gel was characterized by specific physical properties such as organoleptic, pH, spreadability, and adhesivity. This parameter has the function of assessing the quality of the gel in various formulas. Another parameter was to investigate the potential activity of the gel as an anti-inflammatory. The test procedure was performed using the carrageenan-induced rat paw edema model. This acute inflammation model is well-accepted and has long been used to determine its potent anti-inflammatory effect.

## 2.4.1. Organoleptic properties

Gel formulations with or without nutmeg seed extract were evaluated organoleptically for color, texture, and homogeneity. These characteristics were performed by visual observation. The formulation consistency and the particle's coarseness were checked by texture and homogeneity parameters. To assess these parameters, we pressed a small amount of formulated gel between the thumb and the index finger.

## 2.4.2 pH Measurement

Electrochemical pH measurement is used to determine the acidity or alkalinity through a digital pH meter (HANNA HI9813-6 Portable). The principle step was to weigh 1 gram of each gel formula and completely immersed it in the glass electrode. All measurements were taken three times, for which the average reading was recorded [26].

## 2.4.3 Spreadability

The prepared gel spreadability was intended to evaluate the gel's ability to spread and absorb on the skin. This technique was performed by measuring the diameter of spread of 1 gram of gel samples on two horizontal glass slides (10x20cm2) after 60 sec. The standard upper plate for determining gel spreadability was 0.5 g. Each gel formulation was recorded in triplicate [24].

## 2.4.4 Adhesivity

The purpose of gel adhesion testing was to determine the strength that separates the gel from the surface. A tensile adhesion test measures this by placing 2 grams of gel between two resin plates in the designated area. Then clamp the load of 1 kg on the glass plate for 5 minutes. The static resin plate was attached to the adhesion apparatus by applying a trigger load of 80 grams. Further, the evaluation was carried out to determine the length of time using a stopwatch until the resin plate runs out [27].

## 2.4.5 In-vitro anti-inflammatory activity of nutmeg seed extract

The study protocol has obtained ethical approval from Ahmad Dahlan University Ethics Committee with reference number 011502014. The anti-inflammatory properties of nutmeg seed gel products were measured using carrageenan-induce acute inflammation model in hind leg edema of male Wistar rat. The carrageenan-induced paw edema method is a well-defined model of acute inflammation involving various inflammatory mediators, as described by Winter et al. [28]. Initially, healthy Wistar strains (150-200 grams per each) were obtained from an animal breeding house, Faculty of Veterinary Medicine, Gadjah Mada University, Yogyakarta. Rats kept under laboratory conditions were fed twice daily with standard rat chow and water ad libitum. All rats were randomly allocated into six groups: one negative control group, four intervention groups,

and one positive control group. Each group was comprising at least five experimental animals. Before the experiment, the animal had to fast for 24 hours with access to water. 50  $\mu$ l of 1%  $\lambda$ -carrageenan solution (Sigma-Aldrich, Milan, Italy) was suspended in 1% NaCl solution, which had been prepared 1 hour before the experiment. Furthermore, this solution should be injected into the plantar side of the rat's right hind paw. About 0.2 grams of nutmeg seed gel in a different formula was gently applied 50 times with the index finger after 15 minutes of carrageenan injection. The rat paw edema volume was measured immediately using a plethysmometer (model UGO BASILE S.R.L: 7141) at 15 minutes to 3 hours after carrageenan injection. The percentage of inflammation was calculated by the following formula:

% Inflammatory = 
$$\frac{\text{V0 - Vt}}{\text{V0}} \times 100\%$$

**Note**: Vo and Vt are the volumes of rat paw edema from the control group and experimental group.

## 2.5 Data Analysis

The study results were reported descriptively on several test data such as organoleptic properties, pH, adhesiveness, and spreadability. The statistical analysis was performed using One-Way ANOVA and followed by the LSD-post hoc test. A *p*-value less than 0.05 was considered statistically significant.

## 11. RESULTS AND DISCUSSIONS

We have successfully formulated herbal gel from the ethanol extract of nutmeg seed in various formulas. Our study demonstrated that the obtained nutmeg seed gel has a good physical appearance and an anti-inflammatory activity due to acute edema reduction. The detailed finding regarding physical characteristics and anti-inflammatory activity will be presented in the subsection below.

## 3.1 Organoleptic properties

As shown in Table 2, the nutmeg seed gel has a good cosmetic appearance with a soft texture, opaque, brownish-yellow in color, characteristic aromatic odor, and homogeneous without segregation. In general, the dense pigment of nutmeg seed contains more dyes to produce a brownish-yellow gel. Previous studies have also reported that the nutmeg seed gel's physical properties were brownish yellow, fragrant and homogeneous[27][24]. The ethanolic extract does not dissolve completely in the gel bases due to the differences in solubility level.

Table 2. Organoleptic properties of nutmeg seed extract gels

Formula	Physical	Color	Odor	Texture	Homogenity	Segregation
	appearance					
1	Transparant	Clear white	Typical of base	Soft	Homogenous	No
2	Opaque	Brownish yellow	Typical of nutmeg oil	Soft	Homogenous	No
3	Opaque	Brownish yellow	Typical of nutmeg oil	Soft	Homogenous	No
4	Opaque	Brownish yellow	Typical of nutmeg oil	Soft	Homogenous	No
5	Opaque	Brownish yellow	Typical of nutmeg oil	Soft	Homogenous	No

The appearance of nutmeg seed extract gel with various extract concentrations is shown in Figure 1.



Figure 1. The physical appearance of nutmeg seed extract gel at F2 (2%); F3 (4%); F4 (8%) and F5 (12%) (from left to right)

## 3.2 pH Measurement

Each nutmeg seed extract gel formula shows the following pH value F1(4.69  $\pm$  0.74), F2 (5.38  $\pm$  0.78), F3 (5.82  $\pm$  0.15), F4 (6.33  $\pm$  0.13), F5 (6.16  $\pm$  0.24) and F6 (5.87  $\pm$  0.12) (Table 3). These pH value has met the standard criteria for topical skin administration (pH ranged 4-7). pH value measurement could be related to the safety and efficacy of topical drug preparations. Gel acidity and alkalinity will affect skin irritation, penetration and stability of active compounds into the skin. The more acidic the pH value of gel will increase skin irritation, while the more alkaline it will cause dry skin [29][27]. The statistic analysis using the LSD post hoc test showed a significant difference in pH value between F3 to F5 compared to F1 (p<0.001). These results mean that the pH value will increase with the addition of nutmeg extract concentration in the gel base. Meanwhile, the pH value of F1 (gel base) was not significantly different from F2 (extract 2%) (p= 0.121). This defines that both formulas have a similar pH value. As a gelling agent, each end of carbopol's chain has an

acidic carboxylic chemical structure when reacting to water. Carbopol 940 is easily ionized during the neutralization process with the addition of triethanolamine. Therefore, the interaction between carbopol 940 and triethanolamine could affect the pH stability based on the increase in extract concentration [30].

Table 3. The pH value of nutmeg seed extract gel

Formula	pH (Mean $\pm$ SD)		p-value (LSD post hoc test)						
		F1	F2	F3	F4	F5			
F1	4.69±0.74	-	0.121	0.020*	0.002*	0.050*			
F2	5.38±0.78	0.121	-	0.306	0.041*	0.084			
F3	5.82±0.15	0.020*	0.306	-	0.234	0.419			
F4	6.33±0.13	0.002*	0.041*	0.236	-	0.680			
F5	6.16±0.24	0.005*	0.084	0.419	0.680	-			

Note: \* p-value less than 0.05

## 3.3 Adhesivity

Penetration of the active compound gel into the skin will increase along with the enhancement of gel adhesivity properties. The results showed that the average adhesion score of nutmeg seed extract gel was F1 (13.57 $\pm$ 0.11), F2 (18.32 $\pm$ 0.29), F3 (44.33 $\pm$ 0.09), F4 (56.54 $\pm$ 0.19), and F5 (51.12 $\pm$ 0.15) (Table 4). Based on a one-way ANOVA test followed by LSD statistical analysis, the nutmeg extract gel of F4 was significantly greater in adhesion than other formulas (p<0.005). Furthermore, the formulation of F5 with 12% nutmeg seed ethanol extract showed a decrease in adhesiveness compared to F4. This decrease might be due to the pH value being smaller at F5 than F4. The cross-linking between carbopol and other molecules, such as solvents, could also result in ionic attraction, and increasing the gel's viscosity. In acid conditions, increasing the gel's viscosity will have an impact on decreasing the adhesive properties.

Table 4. Adhesive properties of the nutmeg seed extract gel

	Mean $\pm$ SD (second)					
Parameter	F1	F2	F3	F4	F5	1
Adhesivity test (sec)	13.57±0.11	18.32±0.29	44.33±0.09	56.54±0.19	51.12±0.15	0.000

## 3.4 Spreadability

As shown in Table 5, the average spreadability of the nutmeg gel formulations showed F1 (9.38 $\pm$ 0.65), F2 (11.54 $\pm$ 0.21), F3 (13.39 $\pm$ 0.06), F4 (14.85 $\pm$ 0.34), and F5 (19.54 $\pm$ 0.12). These findings concluded that increasing the gel extract concentration would affect the broadest spreadability of the gel. The formula F5 (12%) has the greatest spreadability compared to other formulas. The statistical analysis results through one-way ANOVA stated a significant difference in spreadability between all formulas (p<0.005). Carbopol 940, an acrylic polymer gelling agent, could regulate the viscosity in 1-2% concentrations by producing a three-dimensional matrix to form a viscous gell. The spreadability of the gel formula was highly dependent on the viscosity of the carbopol. The less viscous gelling agent will enhance the spreadability of the gel. The swelling gel process occurs due to the solvent penetration, leading to the cross-linked polymer network to maintain the dosage form and the binding drug particles. However, the alkaline environment pH will increase the density of negative charge, resulting in the gel swollen and facilitating the drug's release [21][30]. Furthermore, the addition of nutmeg seed extract to the gel base will also affect the formation of hydrogen bonds in the physical cross-linking, which influences a reduced viscosity [21].

Table 5. Spreadability properties of the nutmeg seed extract gel

	$Mean \pm SD (cm^2)$					
Parameter	F1	F2	F3	F4	F5	<i>p</i> -value
Spreadability test (cm)	9.38±0.65	11.54±0.21	13.39±0.06	14.85±0.34	19.54±0.12	0.000

## 3.5 In-vitro anti-inflammatory activity of nutmeg seed extract gel

The evaluation of the nutmeg seed herbal gel's anti-inflammatory activity was observed within 180 min (Figure 2). During this time, carrageenan will significantly increase TNF-α, IL-1β, PGE2, iNOS, and COX-2 proteins in peripheral leg inflammation. Carrageenan-induced leg edema follows a model of acute inflammation which consists of two phases: first, which was detected after about 1 hour and was called the fast phase, with the release of histamine and serotonin, and the second stage was called the late phase with the mediators (kinins, prostaglandins) released after 2 and 3 hours, respectively [31]. A previous study suggested that intraplantar carrageenan injection will stimulate rat paw edema and release inflammatory mediators within 180 min [32]. This study is in line with our findings that decreased edema volume also occurred at 180 minutes after intraplantar carrageenan administration.

Figure 2 showed that the formulation of nutmeg seed extract gel has anti-inflammatory effects. It can be proven that the percentage volume of edema in each formula (F2 to F6) was lower than F1 (gel-based negative control). The anti-inflammatory activity of F5 (12%) was higher than that of F2 (2%) and F3 (4%). Furthermore, F4 (8%) and F5 (12%) had anti-inflammatory activity similar to F6 (positive control). This anti-inflammatory activity is influenced by the main compound contained in the nutmeg, namely myristicin. The active compound of nutmeg seed extract, myristicin, has a vital function as an anti-inflammatory by inhibiting chemokines, cytokines, nitrous oxide, and double-stranded growth factors RNA (dsRNA), which are stimulated by macrophages through calcium [33]. Other quercetin compounds also have anti-inflammatory activity by inhibiting the secretion of TNF- $\alpha$ , IL-6, IL-1 $\beta$ , and nitric oxide (NO) [34].

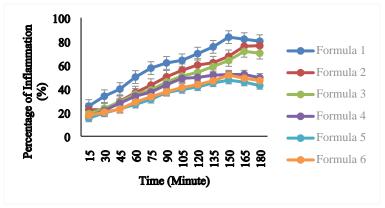


Figure 2. Inflammatory activity of nutmeg seed extract gels.

The volume increase in rat leg edema was used to calculate each formula's anti-inflammatory percentage based on the AUC value. The calculated AUC value and the percentage of anti-inflammatory can be seen in Table 6. Based on the LSD post hoc test analysis reported that the anti-inflammatory activity of F4, F5 did not have a significant difference compared to F6 (positive control). It was concluded that F4 and F5 had similar anti-inflammatory activity as the positive control (1% diclofenac sodium gel in the marketed product). Furthermore, the anti-inflammatory activity of F5 (44.92%) was higher significant difference compared to F4 (37.87%) with p=0.025 The AUC calculation results showed that F5 had the lowest AUC value (5803.72) with the highest percentage of anti-inflammatory activity (44.92%) compared to other formulas (F1 to F5).

Table 6. The percentage of anti-inflammatory activity of nutmeg extract gel was compared between groups.

Formula	AUC	AUC Anti-inflammatory activity (%)	p-value (LSD post hoc test)						
			F1	F2	F3	F4	F5	F6	
F1	10536.94	0	-	0.000*	0.000*	0.000*	0.000*	0.000*	
F2	9304.97	11.69	0.000*	-	0.000*	0.000*	0.000*	0.000*	
F3	7919.26	24.84	0.000*	0.000*	-	0.000*	0.000*	0.000*	
F4	6546.62	37.87	0.000*	0.000*	0.000*	-	0.025*	0.423	
F5	5803.727	44.92	0.000*	0.000*	0.000*	0.025*	-	0.124	
F6	6295.140	40.25	0.000*	0.000*	0.000*	0.423	0.124	-	

Note: \* p-value less than 0.05; F6 is positive control

The highest anti-inflammatory activity of F5 was due to the gel's spreadability and its adhesion to the skin surface. F5 has a good adhesiveness of  $(51.12 \pm 0.15)$  sec with a spreadability of  $(19.54 \pm 0.12)$  cm. The topical drug penetration into the skin was affected by the length of time the gel is in contact with the skin surface. F5 has the highest concentration of nutmeg seed extract (12%) than other formulas (F2 to F4). The high content of myristicin in the F5 gel formula will improve its anti-inflammatory effect by enhancing the drug permeation into the skin. Besides, F5 also had the optimal ternary system composition between ethanol, propylene glycol, and purified water. Propylene glycol (PG) as an enhancer will affect the drug absorption into the skin [35]. A study of topical ibuprofen on human skin reported that ethanol's ternary solvent system, propylene glycol, and water had higher permeability into the skin than the binary solvent system. The ternary system will increase the solubility partition and membrane permeability partitioning by maximizing the active drug flux [36].

## 12. CONCLUSION

These present study indicates that nutmeg gel extract (*Myristica fragrans*, Houtt) with 12% ethanolic extract concentration has a good physical appearance and potent anti-inflammatory activity than other formulas

and marketed product. Hence, nutmeg seed extract could be considered as a potential candidate for antiinflammatory in topical application.

## ACKNOWLEDGEMENTS

The author would like to thank the whole researcher in Ahmad Dahlan University who has supported this research.

## REFERENCES

- [1] S. Kany, J. T. Vollrath, and B. Relja, "Cytokines in inflammatory disease," *Int. J. Mol. Sci.*, vol. 20, no. 23, pp. 1–31, 2019, doi: 10.3390/ijms20236008.
- Y. Su, J. Gao, P. Kaur, and Z. Wang, "Neutrophils and macrophages as targets for development of nanotherapeutics in inflammatory diseases," *Pharmaceutics*, vol. 12, no. 12, pp. 1–24, 2020, doi: 10.3390/pharmaceutics12121222.
- [3] A. L. Loktionov, A. I. Konoplya, M. A. Lunev, and A. V. Karaulov, "Immune and oxidant disorders in the pathogenesis of inflammatory periodontal diseases," *Immunologiya*, vol. 36, no. 5, pp. 319–328, 2015, doi: 10.1111/prd.12002.Inflammatory.
- [4] A. Tedgui and Z. Mallat, "Anti-inflammatory mechanisms in the vascular wall," *Circ. Res.*, vol. 88, no. 9, pp. 877–887, 2001, doi: 10.1161/hh0901.090440.
- [5] M. Del Grossi Moura, L. C. Lopes, M. T. Silva, S. Barberato-Filho, R. H. L. Motta, and C. De Cássia Bergamaschi, "Use of steroid and nonsteroidal anti-inflammatories in the treatment of rheumatoid arthritis: Systematic review protocol," *Med. (United States)*, vol. 97, no. 41, 2018, doi: 10.1097/MD.000000000012658.
- [6] A. Lahoti, B. S. Kalra, and U. Tekur, "Evaluation of the analgesic and anti-inflammatory activity of fixed dose combination: non-steroidal anti-inflammatory drugs in experimental animals.," *Indian J. Dent. Res.*, vol. 25, no. 5, pp. 551–554, 2014, doi: 10.4103/0970-9290.147071.
- [7] K. D. Brandt, "The mechanism of action of nonsteroidal antiinflammatory drugs," *J. Rheumatol.*, vol. 18, no. SUPPL. 27, pp. 120–121, 1991, doi: 10.5772/68090.
- [8] P. Kuropakornpong, A. Itharat, S. Panthong, S. Sireeratawong, and B. Ooraikul, "In Vitro and in Vivo Anti-Inflammatory Activities of Benjakul: A Potential Medicinal Product from Thai Traditional Medicine," *Evidence-based Complement. Altern. Med.*, vol. 2020, 2020, doi: 10.1155/2020/9760948.
- [9] K. R. Patil *et al.*, "Animal models of inflammation for screening of anti-inflammatory drugs: Implications for the discovery and development of phytopharmaceuticals," *Int. J. Mol. Sci.*, vol. 20, no. 18, 2019, doi: 10.3390/ijms20184367.
- [10] S. Tsurufuji, K. Sugio, H. Sato, and K. Ohuchi, "A review of mechanism of action of steroid and non-steroid anti-inflammatory drugs," *Inflamm. Mech. Treat.*, no. 202, pp. 63–78, 1980, doi: 10.1007/978-94-010-9423-8 7.
- [11] V. P. Gullo, J. McAlpine, K. S. Lam, D. Baker, and F. Petersen, "Drug discovery from natural products," *J. Ind. Microbiol. Biotechnol.*, vol. 33, no. 7, pp. 523–531, 2006, doi: 10.1007/s10295-006-0107-2.
- [12] H. Juliani Purba, E. Supriadi Yusufi, and J. Hestina, "Performane and Competitiveness of Indonesian Nutmeg in Export Market," *E3S Web Conf.*, vol. 232, p. 02018, 2021, doi: 10.1051/e3sconf/202123202018.
- [13] J. Marmis, N. S. Tongkeles, Lady Lengkey, and S. Ratulangi, "the Quality of Nutmeg Seeds and Mace," vol. 2, pp. 111–121, 2015.
- [14] K. Karmanah, S. Susanto, W. D. Widodo, and E. Santosa, "The Fruit Characteristics of Ambon Forest Nutmeg (Myristica fatua Houtt) and Banda Nutmeg (Myristica fragrans Houtt)," *J. Ilmu Pertan. Indones.*, vol. 25, no. 2, pp. 292–300, 2020, doi: 10.18343/jipi.25.2.292.
- [15] I. P. S. Kapoor, B. Singh, G. Singh, C. S. De Heluani, M. P. De Lampasona, and C. A. N. Catalan, "Chemical composition and antioxidant activity of essential oil and oleoresins of nutmeg (Myristica fragrans Houtt.) fruits," *Int. J. Food Prop.*, vol. 16, no. 5, pp. 1059–1070, 2013, doi: 10.1080/10942912.2011.576357.
- [16] I. Matulyte *et al.*, "The essential oil and hydrolats from myristica fragrans seeds with magnesium aluminometasilicate as excipient: Antioxidant, antibacterial, and anti-inflammatory activity," *Foods*, vol. 9, no. 1, 2020, doi: 10.3390/foods9010037.
- [17] Jinous Asgarpanah, "Phytochemistry and pharmacologic properties of Myristica fragrans Hoyutt.: A review," *African J. Biotechnol.*, vol. 11, no. 65, 2012, doi: 10.5897/ajb12.1043.
- [18] O. A. Olajide, J. M. Makinde, and S. O. Awe, "Evaluation of the pharmacological properties of nutmeg oil in rats and mice," *Pharm. Biol.*, vol. 38, no. 5, pp. 385–390, 2000, doi: 10.1076/phbi.38.5.385.5976.
- [19] W. K. Zhang et al., "Nutmeg oil alleviates chronic inflammatory pain through inhibition of COX-2

- expression and substance P release in vivo," *Food Nutr. Res.*, vol. 60, no. April, 2016, doi: 10.3402/fnr.v60.30849.
- [20] M. M. Ghareeb, "Design and in vitro characterization of a topical nanoemulsionenriched hydrogel of econazole nitrate," *J. Appl. Pharm. Sci.*, vol. 9, no. 1, pp. 51–57, 2019, doi: 10.7324/JAPS.2019.90108.
- [21] S. A. A. Nabi, M. A. Sheraz, S. Ahmed, N. Mustaan, and I. Ahmad, "Pharmaceutical Gels : A Review," *RADS J. Pharm. Pharm. Sci.*, vol. 4, no. 1, pp. 40–48, 2016.
- [22] C. Tipa, M. T. Cidade, T. Vieira, J. C. Silva, P. I. P. Soares, and J. P. Borges, "Article a new long-term composite drug delivery system based on thermo-responsive hydrogel and nanoclay," *Nanomaterials*, vol. 11, no. 1, pp. 1–22, 2021, doi: 10.3390/nano11010025.
- [23] R. Sánchez-Sánchez *et al.*, "Development of Hydrogel with Anti-Inflammatory Properties Permissive for the Growth of Human Adipose Mesenchymal Stem Cells," *J. Nanomater.*, vol. 2016, 2016, doi: 10.1155/2016/8654937.
- [24] S. Surendran and A. V. Vidyapeetham, "TOPICAL GEL AND ITS IN VITRO EVALUATION FOR ANTINFLAMMATORY Available Online through Research Article," no. April 2016, 2018.
- [25] E. F. Al-Jumaily and M. H. A. Al-Amiry, "Extraction and Purification of Terpenes from Nutmeg (myristica fragrans)," *J. Al-Nahrain Univ. Sci.*, vol. 15, no. 3, pp. 151–160, 2012, doi: 10.22401/jnus.15.3.21.
- [26] S. Karastogianni, S. Girousi, and S. Sotiropoulos, *pH: Principles and Measurement*, 1st ed., no. December. Elsevier Ltd., 2015.
- [27] R. Aiyalu, A. Govindarjan, and A. Ramasamy, "Formulation and evaluation of topical herbal gel for the treatment of arthritis in animal model," *Brazilian J. Pharm. Sci.*, vol. 52, no. 3, pp. 493–507, 2016, doi: 10.1590/s1984-82502016000300015.
- [28] C. A. Winter, E. A. Risley, and G. W. Nuss, "Carrageenin-Induced Edema in Hind Paw of the Rat as an Assay for Antiinflammatory Drugs," *Proc. Soc. Exp. Biol. Med.*, vol. 111, no. 3, pp. 544–547, 1962, doi: 10.3181/00379727-111-27849.
- [29] S. M. Ali and G. Yosipovitch, "Skin pH: From basic science to basic skin care," *Acta Derm. Venereol.*, vol. 93, no. 3, pp. 261–267, 2013, doi: 10.2340/00015555-1531.
- [30] I. Giannopoulou, F. Saïs, and R. Thomopoulos, "Linked data annotation and fusion driven by data quality evaluation," *Rev. des Nouv. Technol. l'Information*, vol. E.28, pp. 257–262, 2015.
- [31] C. W. Li *et al.*, "Anti-inflammatory property of the ethanol extract of the root and rhizome of Pogostemon cablin (Blanco) benth," *Sci. World J.*, vol. 2013, 2013, doi: 10.1155/2013/434151.
- [32] J. C. Fehrenbacher, M. R. Vasko, and D. B. Duarte, "Models of inflammation: carrageenan-or complete freund's adjuvant (CFA)-induced edema and hypersensitivity in the rat," *Curr. Protoc. Pharmacol.*, vol. 56, no. 1, pp. 4–5, 2012.
- [33] J. Y. Lee and W. Park, "Anti-inflammatory effect of myristicin on RAW 264.7 macrophages stimulated with polyinosinic-polycytidylic acid," *Molecules*, vol. 16, no. 8, pp. 7132–7142, 2011, doi: 10.3390/molecules16087132.
- [34] K. Dewi, B. Widyarto, P. Erawijantari, and W. Widowati, "In vitro study of Myristica fragrans seed (Nutmeg) ethanolic extract and quercetin compound as anti-inflammatory agent," *Int. J. Res. Med. Sci.*, vol. 3, no. 9, pp. 2303–2310, 2015, doi: 10.18203/2320-6012.ijrms20150621.
- [35] M. E. Lane, "Skin penetration enhancers," *Int. J. Pharm.*, vol. 447, no. 1–2, pp. 12–21, 2013, doi: 10.1016/j.ijpharm.2013.02.040.
- [36] R. M. Watkinson, C. Herkenne, R. H. Guy, J. Hadgraft, G. Oliveira, and M. E. Lane, "Influence of Ethanol on the Solubility, Ionization and Permeation Characteristics of Ibuprofen in Silicone and Human Skin," *Skin Pharmacol. Physiol.*, vol. 22, no. 1, pp. 15–21, 2009, doi: 10.1159/000183922.

## **BIOGRAPHIES OF AUTHORS (10 PT)**



Azis Iksanudin, is a lecturer in Faculty of Pharmacy, Ahmad Dahlan University, Yogyakarta Indonesia. His research interests are in the field of pharmaceutical technology, natural product, biomaterial, and drug delivery. He has many experience in conduct a researched granted from Higher Education of Indonesia and published some articles in Indonesian and scopus indexed International Journal.



Lolita is a Assistant Professor of Ahmad Dahlan University, Yogyakarta, Indonesia. She is also a Ph.D candidate from Nanjing Medical University, P.R. China. She has published some scientific articles related to social pharmacy, community health in accredited national journal of Indonesia. Her research interest in the topic of clinical pharmacy, pharmacogenomics, community health, and epidemiology. She is also a reviewer in scopus indexed international journals.



**Daniar Dyanas Rais** is an Indonesian pharmacist. She graduated from Bachelor degree programme in Faculty of Pharmacy, Ahmad Dahlan University. Now, she works as a pharmacist in Indonesian state-owned enterprise, Kimia Farma.