

1. PRE-REVIEW OLEH EDITOR



3 AGUSTUS 2024



Participants

Oktira Roka Aji (oktira)

Ichsan Luqmana Indra Putra (ichsanluqmana)

Messages	
Note	From
<p>Dear author,</p> <p>Here's some little correction in your manuscript. I hope we can get the revised manuscript within one week.</p> <p> <a href="#">ichsanluqmana, 11271-Article Text-43387-1-2-20240802.docx</a></p>	<p>ichsanluqmana 2024-08-03 01:42 AM</p>
<p>▶ Dear Editor,</p> <p>I hope this message finds you well.</p> <p>I am writing to inform you that I have completed the revisions based on the feedback and suggestions you provided. I have made the necessary adjustments to address the points that were previously identified.</p> <p>For your convenience, I have attached the updated version of the manuscript to this email. Should you require any further information or additional revisions, please do not hesitate to let me know.</p> <p>Thank you very much for your time and guidance throughout this process. I greatly appreciate your assistance.</p> <p>Best regards,</p> <p>Oktira</p> <p> <a href="#">oktira, 11271-Article Text-43393-1-18-20240803.docx</a></p>	<p>oktira 2024-08-05 05:41 AM</p>

Add Message

## 2. SARAN PERBAIKAN DARI EDITOR

# In Vitro Evaluation of Antioxidant and Lipase Inhibition Properties of Rosella (*Hibiscus sabdariffa* L.) Flower Kombucha

<sup>1\*</sup>Oktira Roka Aji, <sup>2</sup>Anas Tasya Putri, <sup>3</sup>Erisa Budi Handayani, <sup>4</sup>Nisa Ihsani

<sup>1</sup>Department of Biology, Faculty of Applied Science and Technology, Universitas Ahmad Dahlan

<sup>2,3</sup>Microbiology Laboratory, Faculty of Applied Science and Technology, Universitas Ahmad Dahlan

<sup>4</sup>Department of Biotechnology, Faculty of Science and Technology, Universitas Muhammadiyah Bandung

oktira.aji@bio.uad.ac.id\*

\*corresponding author

## ARTICLE INFO

### Article history

Received: August 7<sup>th</sup> 2024

Revised:

Accepted:

### Keywords (Max 5 keywords)

Antioxidant

Fermentation

Kombucha

Lipase Inhibition

Rosella Flower

## ABSTRACT

Kombucha, a traditional fermented beverage made from tea, sugar, and a kombucha culture, is known for its unique flavor, aroma, and health benefits. The fermentation process, typically lasting between 8 and 12 days, influences the beverage's physical and chemical properties. While traditionally brewed from tea, alternative substrates like rosella flowers (*Hibiscus sabdariffa* L.) are also used. Rosella flowers, rich in vitamins and bioactive compounds, have demonstrated potential in combating obesity and exhibiting antioxidant properties. This study aimed to evaluate the chemical and biological characteristics of rosella flower kombucha, particularly its antioxidant and lipase inhibitor activities. The fermentation process led to significant biochemical changes, including a decrease in pH and reducing sugar content, an increase in microbial biomass, and a reduction in ascorbic acid content. The results revealed that rosella flower kombucha had a higher total phenol content and better DPPH radical scavenging activity compared to rosella flower infusion, indicating enhanced antioxidant potential. Additionally, rosella kombucha demonstrated more effective lipase inhibition, suggesting its potential benefits for weight management and metabolic health. These findings emphasize rosella kombucha's potential as a healthful functional beverage, recommending further research in these areas.

This is an open access article under the [CC-BY-SA](#) license.

Commented [I1]: Font?

How to Cite: Last name of first author, abbreviated name., Last name of second author, abbreviated name., & Last name of third author, abbreviated name. (Year of publication of the journal). Full Journal Title. Journal of Biotechnology and Natural Science, 3(1):xx-xx.

## 1. Introduction

Kombucha is a traditional fermented beverage known for its unique flavor and aroma, created from a blend of tea, sugar, and a kombucha culture. Originally introduced by the Chinese, it gained recognition in Indonesia as "kombu tea" during the 1930s. Kombucha, also referred to as Manchurian tea mushroom, is valued for its health benefits due to its diverse beneficial compounds (Naland, 2008). The fermentation process of kombucha typically lasts between 8 and 12 days at temperatures ranging from 18°C to 20°C. Variations in fermentation time—from 7 to 14 days—can significantly alter its physical and chemical properties, including total acidity, pH, alcohol content, and antioxidant levels. Optimal results are achieved with fermentation periods of 5 to 14 days (Sari,



2014). Research indicates that kombucha offers several health benefits, including the ability to inhibit lipid absorption in cases of obesity (Kim et al., 2020).

Susilowati (2013) notes that the type of tea used can influence kombucha fermentation. While traditional kombucha is brewed from tea, alternative substrates are also utilized. One such natural ingredient is rosella flowers (*Hibiscus sabdariffa* L.) (Windyaswari, 2018). Rosella flowers are rich in essential nutrients and chemical compounds, including vitamin C, vitamin A, 18 amino acids, proteins, carbohydrates, thiamine, calcium, anthocyanins, niacin, flavonoids, alkaloids, and stearic acid. The vibrant red color of rosella flowers is attributed to anthocyanins, a type of flavonoid (Gruenwald et al., 2004).

Rosella flowers have demonstrated potential in combating obesity due to their content of flavonoids, anthocyanins, phenolic acids, and organic acids, which are beneficial for pharmacological applications (Nerdy, 2014). Antioxidant activity studies on rosella petals have shown promising results. For example, a study by Nopiyanti and Harjanti (2016) found that ethanol extracts of rosella flowers exhibited an IC<sub>50</sub> of 8.416% in free radical measurements using the DPPH method. Additionally, Ariani et al. (2019) reported that rosella flower ethanol extracts demonstrated a lipase inhibitor activity of 83.65% ± 0.003. Despite the extensive research on various teas as lipase inhibitors and antioxidants, there is a gap in the exploration of rosella kombucha's potential in these areas. Further research is needed to evaluate the effectiveness of rosella kombucha as a lipase inhibitor and antioxidant.

## 2. Methods

### 2.1. Tools and Material

The tools used include analytical scales, test tubes, pipettes, micropipettes, pH meter, beakers, cuvettes, a UV-Vis spectrophotometer, centrifuge tubes, and a vortex mixer. The materials required for the study are dried rosella flowers (*Hibiscus sabdariffa* L.), kombucha starter culture, SCOBY (Symbiotic Culture of Bacteria and Yeast), gallic acid, DPPH (1,1-Diphenyl-2-picrylhydrazyl) solution, pancreatic lipase, buffer Tris-HCl, pH 8, pNPP (p-Nitrophenyl Palmitate), acetonitrile, and DMSO.

### 2.2. Kombucha Starter Preparation

Kombucha starter culture known as "SCOBY", (generally consisting of *Acetobacter xylinum*, *Gluconobacter* and *Saccharomyces cerevisiae*) obtained from Laboratory of Microbiology Universitas Ahmad Dahlan, Yogyakarta. The starter culture used in this study was stored in a refrigerator (4°C) and consisted of broth acid and a layer of cellulose ("tea mold" floating on the surface).

### 2.3. Rosella Flower Kombucha Preparation

The process of making rosella flower kombucha began with the creation of a rosella flower infusion. This involved steeping 1 gram of dried rosella flowers in 200 mL of boiling water for 15 minutes. Following the brewing process, the resulting mixture was strained into a sterile glass container or jar to extract the liquid filtrate. Subsequently, 20 grams of powdered sugar, constituting a 10% weight/volume ratio, were added to the filtrate and stirred until the sugar had completely dissolved. The sweetened infusion was then covered and allowed to cool to approximately 30°C. At this lower temperature, 10% of the starter culture, consisting of 12 mL of liquid starter and 8 grams of SCOBY starter material, was introduced. The jar was securely covered with sterile gauze and the mixture was left to ferment at room temperature (28-30°C). The kombucha was fermented for various durations (0, 3, 6, 9, and 12 days).

### 2.4. Chemical Characteristics of Kombucha

The chemical characteristics of kombucha were assessed through pH measurement, reducing sugar content, total phenol content, and vitamin C (ascorbic acid) content. The pH acidity of the product was measured using a pH meter with three replicates. Reducing sugar content was determined using the Somogyi-Nelson method. The total phenolic content was evaluated following the Folin-Ciocalteu method described by Agbor et al. (2014). For the vitamin C content test, the iodimetric titration method was employed. Twenty-five mL of rosella flower kombucha was pipetted and diluted with 100 mL of distilled water, and the initial weight was recorded. Ten mL of this diluted sample was transferred into a 250 mL Erlenmeyer flask, and 2 mL of 1% amylum indicator

solution was added. Titration was then performed using 0.01 N iodine until a blue color developed. The concentration of vitamin C was calculated using the formula:  $(A \times 0.88 \times FP) / W$ , where A represents the volume of iodine used for titration (mL), 0.88 mg is the amount of ascorbic acid equivalent to 1 mL of 0.01 N iodine solution, FP is the dilution factor, and W is the sample weight (mg).

## 2.5. Biological Characteristics of Kombucha

The biological characteristics of kombucha were assessed by measuring the mushroom dry weight and optical density. The optical density (OD) of the kombucha was measured using a spectrophotometer at a wavelength of 600 nm. The dry weight of the SCOBY, also referred to as mushroom dry weight, was determined by drying it in an oven at 80°C until a constant weight was reached, after which it was weighed.

## 2.6. Antioxidant Activity Assay Using the DPPH Method

The sample tested for antioxidant activity was an infusion of rosella flowers and kombucha that had been fermented for 12 days. The antioxidant activity of rosella flower kombucha was evaluated using the DPPH method. Initially, a DPPH solution was prepared by dissolving 0.5 mg of DPPH in 50 mL of methanol p.a to achieve a 10 mg/L concentration. The control solution was made by homogenizing this mother solution and incubating it for 30 minutes at 37°C. For the antioxidant activity test, 1 mL of the 10 mg/L DPPH solution was combined with 50 µL of the sample liquid, and the mixture was diluted with methanol p.a to a final volume of 5 mL. This mixture was then incubated for 30 minutes at 37°C, after which the absorbance was measured at 515 nm. Antioxidant activity was assessed based on the inhibition of DPPH radical uptake. The percentage inhibition was calculated using the formula:

$$\% \text{ DPPH Inhibition} = \frac{\text{Abs Blanko} - \text{Abs sampel}}{\text{Abs Blanko}} \times 100\%$$

where Abs<sub>Blank</sub> denotes the absorbance of the DPPH radicals at 515 nm, and Abs<sub>Sample</sub> represents the absorbance of the sample at the same wavelength.

## 2.7. In Vitro Lipase Inhibitor Activity Assay

The samples tested for *in vitro* lipase inhibitor activity assay included rosella flower infusion and kombucha, which had been fermented for 12 days. The lipase inhibitor activity test was conducted as follows: A stock solution of rosella flower infusion was first prepared by taking 100 mL of the pure infusion. Kombucha samples that had undergone fermentation for 3, 6, 9, and 12 days were then centrifuged to obtain clear solutions, which were subsequently stored at 4°C. For the enzyme preparation, a 1 mg/mL stock solution of pancreatic lipase was prepared by dissolving 10 mg of the enzyme in 10 mL of a 50 mM Tris-HCl buffer with a pH of 8. Additionally, a 50 mM stock solution of pNPP in acetonitrile was prepared by dissolving 0.19 g of pNPP in 9.81 mL of acetonitrile, making up a final volume of 10 mL. To test the inhibitor activity, a mixture was prepared containing 0.1 mL of the 1 mg/mL lipase solution, 0.2 mL of either the rosella flower infusion or kombucha (at various concentrations), and 0.7 mL of the 50 mM Tris-HCl buffer (pH 8). This mixture was incubated for 15 minutes at room temperature (37°C), after which 0.1 mL of the 50 mM pNPP solution was added. The solution was then incubated for an additional 30 minutes at room temperature, and the absorbance was measured at 410 nm using a spectrophotometer. The test was conducted in duplicate. For the positive control test, a stock solution of orlistat was prepared by dissolving 20 mL of orlistat in 20 mL of distilled water. The enzyme stock solution and the pNPP substrate were prepared in the same manner as previously described. The activity test involved mixing 0.1 mL of the 1 mg/mL lipase solution with 0.2 mL of orlistat (at various concentrations) and 0.7 mL of the 50 mM Tris-HCl buffer (pH 8). This mixture was incubated for 15 minutes at 37°C, followed by the addition of 0.1 mL of the 50 mM pNPP solution. After another 30 minutes of incubation at room temperature, the absorbance was measured at 410 nm using a spectrophotometer. This test was also performed in duplicate. The percentage of inhibition was calculated using the formula:

$$\% \text{ Inhibition} = \frac{A_0 - A}{A_0} \times 100\%$$

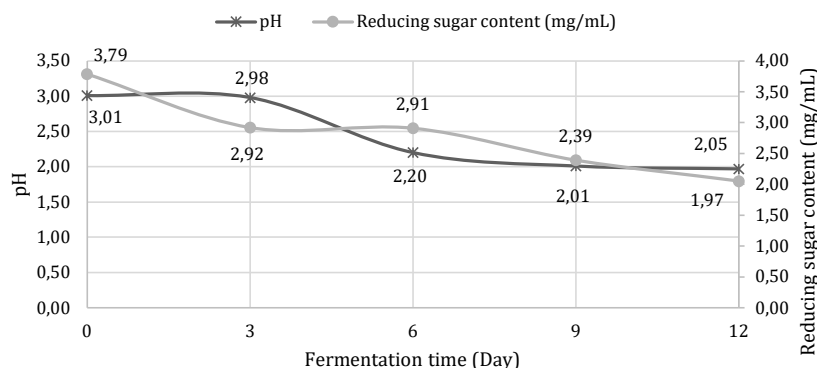
where  $A_0$  represented the lipase activity without an inhibitor, and A represented the activity in the presence of an inhibitor.

Commented [I2]: Citation of this formula?

Commented [I3]: Citation of the formula

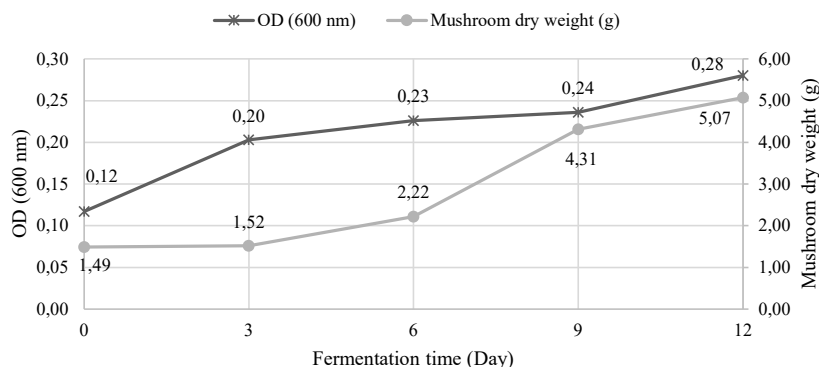
### 3. Results and Discussion

During the 12-day fermentation process of roselle flower kombucha, significant biochemical changes occur that affect the final product's quality and nutritional value. As showed in Figure 1, the pH of the kombucha decreases from 3.79 to 1.97, indicating a rise in acidity. This acidification, driven by microbial activity, aligns with findings from previous studies by Wang et al. (2023) demonstrated that the pH in kombucha typically drops as fermentation progresses due to the production of acetic acid, gluconic acid, and glucuronic acid by the kombucha symbiotic culture of bacteria and yeast (SCOBY). Concurrent with the decrease in pH, the reducing sugar content drops from 3.01 mg/mL to 1.97 mg/mL. This gradual decline in sugar levels, in line with observations by Kushargina et al. (2024), suggests that sugars are being metabolized by the microbes, leading to acid production and contributing to the pH reduction. The study by Cohen et al. (2023) noted a similar trend, where a decrease in reducing sugars was closely associated with the formation of organic acids throughout the fermentation process.



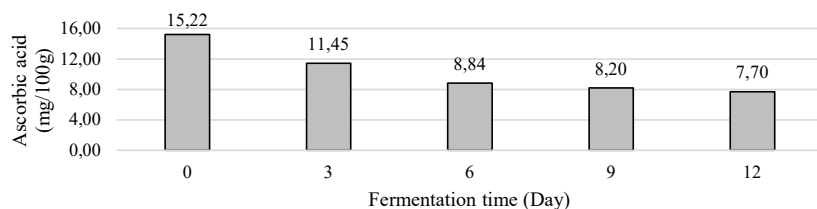
**Figure 1.** Changes in pH and reducing sugar content during rosella flower kombucha fermentation

Changes in OD<sub>600</sub> and dry weight of the mushroom (cellulose pellicle) during 12 days of roselle flower kombucha fermentation are depicted in Figure 2. The optical density (OD) at 600 nm, a measure of microbial growth, increases from 0.12 to 0.28, indicating a steady accumulation of microbial biomass. This finding is consistent with previous studies, such as those by Ahmed et al. (2020), which reported an increase in OD<sub>600</sub> corresponding with the growth of the kombucha SCOBY throughout fermentation. This microbial proliferation is correlated with a substantial increase in the mushroom dry weight, from 1.49 grams to 5.07 grams, with the most significant growth occurring after day 6. This growth pattern supports the findings of Charoenrak et al. (2023), who observed similar trends in cellulose pellicle development during kombucha fermentation. The term "mushroom" here refers to the cellulose pellicle produced by a symbiotic culture of bacteria and yeast (SCOBY), including species such as *Acetobacter xylinum*, known for its cellulose-synthesizing abilities. This suggests that the conditions created during the early stages of fermentation are favorable for microbial growth, which in turn supports the accumulation of mushroom biomass, likely through the activities of these bacteria and yeast in breaking down substrates that facilitate pellicle development.



**Figure 2.** Changes in optical density (OD) and mushroom dry weight during rosella flower kombucha fermentation

In relation to the ascorbic acid content, Figure 3 shows a marked decline from 15.22 mg/100g to 7.70 mg/100g over the fermentation period. The most rapid reduction occurs in the early stages (days 0 to 3), after which the decline slows and approaches a more stable value by day 12. This reduction in ascorbic acid could be attributed to its degradation under acidic conditions or microbial utilization. Vitamin C, an essential vitamin but unstable molecule, is prone to degradation due to its sensitivity to light, alkaline pH, high temperatures, oxygen, heavy metals, UV rays, and extended storage times. During fermentation, this degradation is exacerbated by the acidic conditions and the microbial activities involved. Specifically, the oxidation process and the presence of ascorbate oxidase enzymes produced by the fermenting microorganisms contribute significantly to the loss of ascorbic acid (Jafarpour & Hashemi, 2023).



**Figure 3.** Ascorbic acid content over time during rosella flower kombucha fermentation

The comparison between Roselle flower infusion and Roselle kombucha reveals several notable differences in their chemical and antioxidant properties (Table 1.). Roselle kombucha has a higher total phenol content (0.02  $\mu\text{L/mL}$ ) compared to Roselle flower infusion (0.01  $\mu\text{L/mL}$ ), suggesting that kombucha may offer a more substantial antioxidant potential due to its increased concentration of phenolic compounds. This is further supported by the higher % inhibition of DPPH at 25% (v/v) observed in Roselle kombucha (62.5%) compared to the Roselle flower infusion (55.88%), indicating that kombucha is more effective at scavenging free radicals. Additionally, the IC<sub>50</sub> value for antioxidant activity, which reflects the concentration required to inhibit 50% of free radicals, is lower in Roselle kombucha (16.25%) than in Roselle flower infusion (16.85%), demonstrating that kombucha is slightly more effective as an antioxidant. Furthermore, when assessing lipase inhibition, Roselle kombucha exhibits a lower IC<sub>50</sub> (0.147%) than Roselle flower infusion (0.253%), suggesting that kombucha is more effective in inhibiting lipase activity. The results of this study are in line with butterfly pea flower kombucha which is more effective in inhibiting pancreatic lipase in vitro than butterfly pea flower tea infusion (Aji et al., 2023). These



findings also align with previous research indicating that kombucha, often fermented with various herbs and fruits, can have enhanced antioxidant and enzyme-inhibiting properties compared to its non-fermented counterparts. Studies such as those by Cardoso et al. (2020) and Jayabalan and Waisundara (2019) have shown that kombucha fermentation can increase the concentration of bioactive compounds and improve antioxidant capacities.

Table 1. Comparison of total phenol content, antioxidant activity, and in vitro lipase inhibition between roselle flower infusion and roselle flower kombucha

	Roselle flower infusion	Roselle flower kombucha (12 <sup>th</sup> day fermentation)
Total phenol content (GAE $\mu$ L/mL)	0,01	0,02
% inhibition of DPPH at 25% (v/v)	55,88	62,5
IC50 antioxidant activity (% v/v)	16,85	16,25
IC50 inhibition of lipase activity (% v/v)	0,253	0,147

The study comparing Roselle flower infusion and Roselle kombucha highlights several significant implications for health and functional beverage development. Roselle kombucha's superior antioxidant properties, evidenced by its higher total phenol content and better DPPH inhibition, suggest it could offer more effective protection against oxidative stress compared to Roselle flower infusion. This enhanced antioxidant activity may contribute to reduced risk of chronic diseases and support overall health. Additionally, kombucha's greater lipase inhibition suggests potential benefits for managing fat digestion and absorption, making it a promising option for weight management and metabolic health.

#### 4. Conclusion

The study demonstrates that rosella flower kombucha possesses superior antioxidant and in vitro lipase inhibitory properties compared to rosella flower infusion, likely due to the fermentation process that increases the concentration of bioactive compounds. The findings suggest that kombucha, particularly when fermented with functional ingredients like rosella, could serve as a potent health-promoting beverage. The enhanced antioxidant activity and lipase inhibition indicate potential benefits in managing oxidative stress and fat metabolism, making rosella kombucha a promising candidate for functional food development.

#### REFERENCES

- Agbor, G. A., Vinson, J. A., & Donnelly, P. E. (2014). Folin-Ciocalteu reagent for polyphenolic assay. *International Journal of Food Science, Nutrition and Dietetics (IJFS)*, 3, 1-10. <https://doi.org/10.19070/2326-3350-1400028>
- Ahmed, S., Mohamed, S., Hikal, M., Khadiga, A., & Abou-Taleb, M. (2020). Biological, chemical and antioxidant activities of different types Kombucha. *Annals of Agricultural Sciences*, 65, 35-44. <https://doi.org/10.1016/j.aos.2020.04.001>
- Aji, O. R., Putri, N. F., & Putri, D. A. (2023). Pancreatic lipase inhibitory activity of butterfly pea flower (*Clitoria ternatea*) Kombucha. *Biogenesis Jurnal Ilmiah Biologi*, 11(2), 151-159. <https://doi.org/10.24252/bio.v11i2.35724>
- Ariani, L. W., Ungsari, R. E. P., & Mighfar, S. (2019). Formulasi mikropartikel ekstrak etanol bunga rosela (*Hibiscus Sabdariffa* L.) dalam sediaan sirup dan potensinya sebagai obat herbal antiobesitas. *Jurnal Ilmiah Cendekia Eksakta*, 4, 1.
- Cardoso, R. R., Neto, R. O., D'Almeida, C. T. D. S., Nascimento, T. P. D., Pressete, C. G., Azevedo, L., Martino, H. S. D., Cameron, L. C., Ferreira, M. S. L., & De Barros, F. A. R. (2020). Kombuchas from green and black teas have different phenolic profiles, which impacts their antioxidant capacities, antibacterial and antiproliferative activities. *Food Research International*, 128, 108782. <https://doi.org/10.1016/j.foodres.2019.108782>

**Commented [I4]:** Beware in the scientific name, there are some  
scientific name didn't write in italic

Charoenrak, S., Charumane, S., Sirisa-Ard, P., Bovonsombut, S., Kumdhithahutsawakul, L., Kiatkarun, S., Pathom-Aree, W., Chitov, T., & Bovonsombut, S. (2023). Nanobacterial cellulose from Kombucha fermentation as a potential protective carrier of *Lactobacillus plantarum* under simulated gastrointestinal tract conditions. *Polymers*, 15(6), 1356. <https://doi.org/10.3390/polym15061356>

Cohen, G., Sela, D. A., & Nolden, A. A. (2023). Sucrose concentration and fermentation temperature impact the sensory characteristics and liking of kombucha. *Foods*, 12(16), 3116. <https://doi.org/10.3390/foods12163116>

Gruenewald, J., Brendler, T., & Jaenicke, C. (2004). PDR for herbal medicines, third edition. Medical Economics Company, New Jersey.

Jafarpour, D., & Hashemi, S. M. B. (2023). Pure and co-fermentation of quinoa seeds by *Limosilactobacillus fermentum* and *Lactocaseibacillus rhamnosus*: Bioactive content, antidiabetic and antioxidant activities. *Fermentation*, 9(2), 80. <https://doi.org/10.3390/fermentation9020080>

Jayabalan, R., Malbaša, R. V., Lončar, E. S., Vitas, J. S., & Sathishkumar, M. (2014). A review on kombucha tea: Microbiology, composition, fermentation, beneficial effects, toxicity, and tea fungus. *Comprehensive Reviews in Food Science and Food Safety*, 13, 538-550. <https://doi.org/10.1111/1541-4337.12073>

Kim, D. H., Park, Y. H., Lee, J. S., Jeong, H. I., Lee, K. W., & Kang, T. H. (2020). Antiobesity effect of DKB-117 through the inhibition of pancreatic lipase and  $\alpha$ -amylase activity. *Nutrients*, 12, 3053. <https://doi.org/10.3390/nu12103053>

Kushargina, R., Rimbawan, R., Dewi, M., & Damayanthi, E. (2024). Effect of sugar addition and fermentation process on flavonoids, phenols, and antioxidant activity of Telang (*Clitoria ternatea* L.) kombucha. *Tropical Journal of Natural Product Research*, 8(3). <https://doi.org/10.26538/tjnpr/v8i3.39>

Naland, P. (2008). Kombucha teh dengan seribu khasiat. Agromedia Pustaka, Jakarta.

Nerdy, N. (2014). In silico docking of chemical compounds from roselle calyces (*Hibiscus Sabdariffa* L.) as antidiabetic. *International Journal of ChemTech Research*, 7, 148-152.

Nopiyantri, D., & Harjanti, A. (2016). Analisis stabilitas senyawa aktif antioksidan kelopak bunga rosella (*Hibiscus sabdariffa* L.) pada penggunaannya sebagai bahan tambahan pangan alami. *Jurnal Farmasi Indonesia*, 13, 101-110.

Sari, P. (2014). Perbandingan aktivitas antioksidan kombucha teh hijau (*Camelia sinensis*) dengan teh daun mangga (*Mangifera indica*) dipengaruhi oleh lama fermentasi. Skripsi. Surakarta: Universitas Muhammadiyah Surakarta.

Susilowati, T. (2013). Perbedaan waktu fermentasi dalam pembuatan teh kombucha dari ekstrak teh hijau lokal Arraca kiara, Arraca yabukita, Pekoe dan Dewata sebagai minuman fungsional untuk antioksidan. Prosiding SNST ke-4. Semarang: Universitas Wahid Hasyim.

Wang, B., Rutherford-Markwick, K., Naren, N., Zhang, X., & Mutukumira, A. N. (2023). Microbiological and physico-chemical characteristics of black tea kombucha fermented with a New Zealand starter culture. *Foods*, 12(12), 2314. <https://doi.org/10.3390/foods12122314>

Windyaswari, A. (2018). Pengaruh teknik dan pelarut ekstraksi terhadap aktivitas antioksidan dan empat jenis ekstrak daun rosella (*Hibiscus Sabdariffa* L.). *TM Conference Series*, 1, 14-1. <https://doi.org/10.32734/tm.v1i3.254>

### 3. PERMINTAAN REVISI

19 SEPTEMBER 2024

---

## [JBNS] Editor Decision

1 message

---

**Ichsan Luqmana Indra Putra** <ichsan.luqmana@bio.uad.ac.id>

Thu, Sep 19, 2024 at 2:12 PM

To: Oktira Roka Aji <oktira.aji@bio.uad.ac.id>, Anas Tasya Putri <anas1800017104@webmail.uad.ac.id>, Erisa Budi Handayani <erisa1800017135@webmail.uad.ac.id>, Nisa Ihsani <nisaihsani@umbandung.ac.id>

Oktira Roka Aji, Anas Tasya Putri, Erisa Budi Handayani, Nisa Ihsani:

We have reached a decision regarding your submission to Journal of Biotechnology and Natural Science, "In Vitro Evaluation of Antioxidant and Lipase Inhibition Properties of Rosella (Hibiscus sabdariffa L.) Flower Kombucha".

Our decision is: Revisions Required

---

### 2 attachments



**C-11271-44303-1-5-20240903.docx**

82K



**C-11271-44301-1-5-20240903.docx**

93K

#### 4. SARAN PERBAIKAN DARI REVIEWER

# In Vitro Evaluation of Antioxidant and Lipase Inhibition Properties of Rosella (*Hibiscus sabdariffa* L.) Flower Kombucha

## ARTICLE INFO

### Article history

Received: August 7<sup>th</sup> 2024  
Revised:  
Accepted:

### Keywords

Antioxidant  
Fermentation  
Kombucha  
Lipase Inhibition  
Rosella Flower

## ABSTRACT

Kombucha, a traditional fermented beverage made from tea, sugar, and a kombucha culture, is known for its unique flavor, aroma, and health benefits. The fermentation process, typically lasting between 8 and 12 days, influences the beverage's physical and chemical properties. While traditionally brewed from tea, alternative substrates like rosella flowers (*Hibiscus sabdariffa* L.) are also used. Rosella flowers, rich in vitamins and bioactive compounds, have demonstrated potential in combating obesity and exhibiting antioxidant properties. This study aimed to evaluate the chemical and biological characteristics of rosella flower kombucha, particularly its antioxidant and lipase inhibitor activities. The fermentation process led to significant biochemical changes, including a decrease in pH and reducing sugar content, an increase in microbial biomass, and a reduction in ascorbic acid content. The results revealed that rosella flower kombucha had a higher total phenol content and better DPPH radical scavenging activity compared to rosella flower infusion, indicating enhanced antioxidant potential. Additionally, rosella kombucha demonstrated more effective lipase inhibition, suggesting its potential benefits for weight management and metabolic health. These findings emphasize rosella kombucha's potential as a healthful functional beverage, recommending further research in these areas.

This is an open access article under the [CC-BY-SA](#) license.

How to Cite: Last name of first author, abbreviated name., Last name of second author, abbreviated name., & Last name of third author, abbreviated name. (Year of publication of the journal). Full Journal Title. Journal of Biotechnology and Natural Science, 3(1):xx-xx.

## 1. Introduction

Kombucha is a traditional fermented beverage known for its unique flavor and aroma, created from a blend of tea, sugar, and a kombucha culture. Originally introduced by the Chinese, it gained recognition in Indonesia as "kombu tea" during the 1930s. Kombucha, also referred to as Manchurian tea mushroom, is valued for its health benefits due to its diverse beneficial compounds (Naland, 2008). The fermentation process of kombucha typically lasts between 8 and 12 days at temperatures ranging from 18°C to 20°C. Variations in fermentation time—from 7 to 14 days—can significantly alter its physical and chemical properties, including total acidity, pH, alcohol content, and antioxidant levels. Optimal results are achieved with fermentation periods of 5 to 14 days (Sari, 2014). Research indicates that kombucha offers several health benefits, including the ability to inhibit lipid absorption in cases of obesity (Kim et al., 2020).

Susilowati (2013) notes that the type of tea used can influence kombucha fermentation. While traditional kombucha is brewed from tea, alternative substrates are also utilized. One such natural ingredient is rosella flowers (*Hibiscus sabdariffa* L.) (Windyaswari, 2018). Rosella flowers are rich in essential nutrients and chemical compounds, including vitamin C, vitamin A, 18 amino acids,

**Commented [A1]:** Authors can add the reason to innovate and utilize Rosella flower for kombucha making.



proteins, carbohydrates, thiamine, calcium, anthocyanins, niacin, flavonoids, alkaloids, and stearic acid. The vibrant red color of rosella flowers is attributed to anthocyanins, a type of flavonoid (Gruenwald et al., 2004).

Rosella flowers have demonstrated potential in combating obesity due to their content of flavonoids, anthocyanins, phenolic acids, and organic acids, which are beneficial for pharmacological applications (Nerdy, 2014). Antioxidant activity studies on rosella petals have shown promising results. For example, a study by Nopiyanti and Harjanti (2016) found that ethanol extracts of rosella flowers exhibited an  $IC_{50}$  of 8.416% in free radical measurements using the DPPH method. Additionally, Ariani et al. (2019) reported that rosella flower ethanol extracts demonstrated a lipase inhibitor activity of  $83.65\% \pm 0.003$ . Despite the extensive research on various teas as lipase inhibitors and antioxidants, there is a gap in the exploration of rosella kombucha's potential in these areas. Further research is needed to evaluate the effectiveness of rosella kombucha as a lipase inhibitor and antioxidant.

## 2. Methods

### 2.1. Tools and Material

The tools used include analytical scales, test tubes, pipettes, micropipettes, pH meter, beakers, cuvettes, a UV-Vis spectrophotometer, centrifuge tubes, and a vortex mixer. The materials required for the study are dried rosella flowers (*Hibiscus sabdariffa* L.), kombucha starter culture, SCOBY (Symbiotic Culture of Bacteria and Yeast), gallic acid, DPPH (1,1-Diphenyl-2-picrylhydrazyl) solution, pancreatic lipase, buffer Tris-HCl, pH 8, pNPP (p-Nitrophenyl Palmitate), acetonitrile, and DMSO.

### 2.2. Kombucha Starter Preparation

Kombucha starter culture known as "SCOBY", (generally consisting of *Acetobacter xylinum*, *Gluconobacter* and *Saccharomyces cerevisiae*) obtained from Laboratory of Microbiology Universitas Ahmad Dahlan, Yogyakarta. The starter culture used in this study was stored in a refrigerator (4°C) and consisted of broth acid and a layer of cellulose ("tea mold" floating on the surface).

### 2.3. Rosella Flower Kombucha Preparation

The process of making rosella flower kombucha began with the creation of a rosella flower infusion. This involved steeping 1 gram of dried rosella flowers in 200 mL of boiling water for 15 minutes. Following the brewing process, the resulting mixture was strained into a sterile glass container or jar to extract the liquid filtrate. Subsequently, 20 grams of powdered sugar, constituting a 10% weight/volume ratio, were added to the filtrate and stirred until the sugar had completely dissolved. The sweetened infusion was then covered and allowed to cool to approximately 30°C. At this lower temperature, 10% of the starter culture, consisting of 12 mL of liquid starter and 8 grams of SCOBY starter material, was introduced. The jar was securely covered with sterile gauze and the mixture was left to ferment at room temperature (28-30°C). The kombucha was fermented for various durations (0, 3, 6, 9, and 12 days).

### 2.4. Chemical Characteristics of Kombucha

The chemical characteristics of kombucha were assessed through pH measurement, reducing sugar content, total phenol content, and vitamin C (ascorbic acid) content. The pH acidity of the product was measured using a pH meter with three replicates. Reducing sugar content was determined using the Somogyi-Nelson method. The total phenolic content was evaluated following the Folin-Ciocalteu method described by Agbor et al. (2014). For the vitamin C content test, the iodimetric titration method was employed. Twenty-five mL of rosella flower kombucha was pipetted and diluted with 100 mL of distilled water, and the initial weight was recorded. Ten mL of this diluted sample was transferred into a 250 mL Erlenmeyer flask, and 2 mL of 1% amylum indicator solution was added. Titration was then performed using 0.01 N iodine until a blue color developed. The concentration of vitamin C was calculated using the formula:  $(A \times 0.88 \times FP) / W$ , where A represents the volume of iodine used for titration (mL), 0.88 mg is the amount of ascorbic acid equivalent to 1 mL of 0.01 N iodine solution, FP is the dilution factor, and W is the sample weight (mg).

### 2.5. Biological Characteristics of Kombucha

The biological characteristics of kombucha were assessed by measuring the mushroom dry weight and optical density. The optical density (OD) of the kombucha was measured using a spectrophotometer at a wavelength of 600 nm. The dry weight of the SCOBY, also referred to as mushroom dry weight, was determined by drying it in an oven at 80°C until a constant weight was reached, after which it was weighed.

#### 2.6. Antioxidant Activity Assay Using the DPPH Method

The sample tested for antioxidant activity was an infusion of rosella flowers and kombucha that had been fermented for 12 days. The antioxidant activity of rosella flower kombucha was evaluated using the DPPH method. Initially, a DPPH solution was prepared by dissolving 0.5 mg of DPPH in 50 mL of methanol p.a to achieve a 10 mg/L concentration. The control solution was made by homogenizing this mother solution and incubating it for 30 minutes at 37°C. For the antioxidant activity test, 1 mL of the 10 mg/L DPPH solution was combined with 50 µL of the sample liquid, and the mixture was diluted with methanol p.a to a final volume of 5 mL. This mixture was then incubated for 30 minutes at 37°C, after which the absorbance was measured at 515 nm. Antioxidant activity was assessed based on the inhibition of DPPH radical uptake. The percentage inhibition was calculated using the formula (Aji et al., 2023a):

$$\% \text{ DPPH Inhibition} = \frac{\text{Abs}_{\text{Blanko}} - \text{Abs}_{\text{sample}}}{\text{Abs}_{\text{Blanko}}} \times 100\%$$

where Abs<sub>Blank</sub> denotes the absorbance of the DPPH radicals at 515 nm, and Abs<sub>Sample</sub> represents the absorbance of the sample at the same wavelength.

#### 2.7. In Vitro Lipase Inhibitor Activity Assay

The samples tested for *in vitro* lipase inhibitor activity assay included rosella flower infusion and kombucha, which had been fermented for 12 days. The lipase inhibitor activity test was conducted as follows: A stock solution of rosella flower infusion was first prepared by taking 100 mL of the pure infusion. Kombucha samples that had undergone fermentation for 3, 6, 9, and 12 days were then centrifuged to obtain clear solutions, which were subsequently stored at 4°C. For the enzyme preparation, a 1 mg/mL stock solution of pancreatic lipase was prepared by dissolving 10 mg of the enzyme in 10 mL of a 50 mM Tris-HCl buffer with a pH of 8. Additionally, a 50 mM stock solution of pNPP in acetonitrile was prepared by dissolving 0.19 g of pNPP in 9.81 mL of acetonitrile, making up a final volume of 10 mL. To test the inhibitor activity, a mixture was prepared containing 0.1 mL of the 1 mg/mL lipase solution, 0.2 mL of either the rosella flower infusion or kombucha (at various concentrations), and 0.7 mL of the 50 mM Tris-HCl buffer (pH 8). This mixture was incubated for 15 minutes at room temperature (37°C), after which 0.1 mL of the 50 mM pNPP solution was added. The solution was then incubated for an additional 30 minutes at room temperature, and the absorbance was measured at 410 nm using a spectrophotometer. The test was conducted in duplicate. For the positive control test, a stock solution of orlistat was prepared by dissolving 20 mL of orlistat in 20 mL of distilled water. The enzyme stock solution and the pNPP substrate were prepared in the same manner as previously described. The activity test involved mixing 0.1 mL of the 1 mg/mL lipase solution with 0.2 mL of orlistat (at various concentrations) and 0.7 mL of the 50 mM Tris-HCl buffer (pH 8). This mixture was incubated for 15 minutes at 37°C, followed by the addition of 0.1 mL of the 50 mM pNPP solution. After another 30 minutes of incubation at room temperature, the absorbance was measured at 410 nm using a spectrophotometer. This test was also performed in duplicate. The percentage of inhibition was calculated using the formula (Aji et al., 2023b):

$$\% \text{ Inhibition} = \frac{A_0 - A}{A_0} \times 100\%$$

where A<sub>0</sub> represented the lipase activity without an inhibitor, and A represented the activity in the presence of an inhibitor.

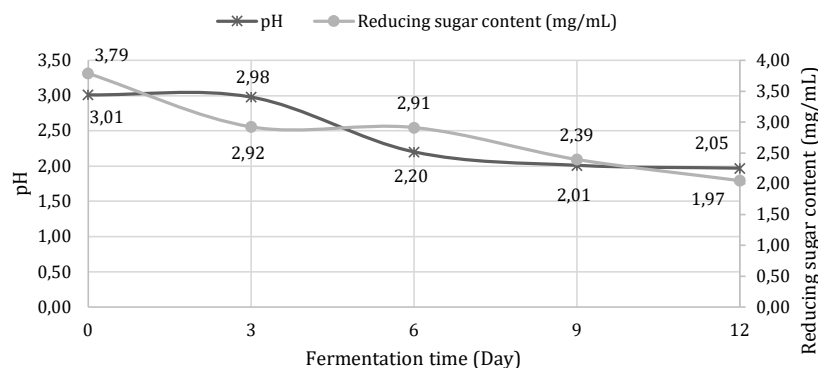
### 3. Results and Discussion

During the 12-day fermentation process of roselle flower kombucha, significant biochemical changes occur that affect the final product's quality and nutritional value. As showed in Figure 1, the pH of the kombucha decreases from 3.79 to 1.97, indicating a rise in acidity. This acidification, driven by microbial activity, aligns with findings from previous studies by Wang et al. (2023) demonstrated that the pH in kombucha typically drops as fermentation progresses due to the production of acetic acid, gluconic acid, and glucuronic acid by the kombucha symbiotic culture of bacteria and yeast

Commented [A2]: The correct term is Pellicle.

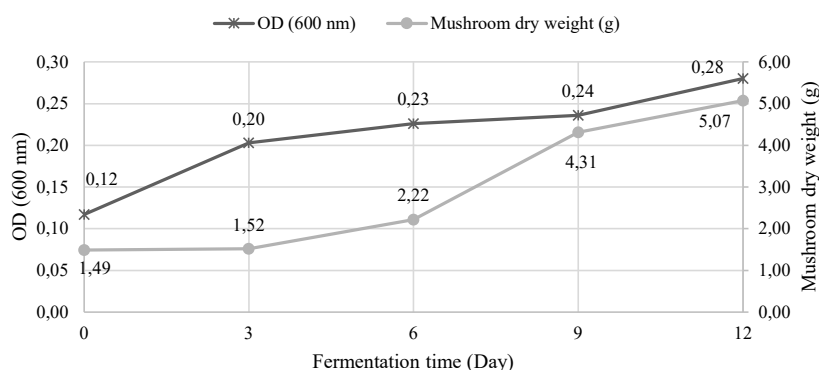


(SCOBY). Concurrent with the decrease in pH, the reducing sugar content drops from 3.01 mg/mL to 1.97 mg/mL. This gradual decline in sugar levels, in line with observations by Kushargina et al. (2024), suggests that sugars are being metabolized by the microbes, leading to acid production and contributing to the pH reduction. The study by Cohen et al. (2023) noted a similar trend, where a decrease in reducing sugars was closely associated with the formation of organic acids throughout the fermentation process.



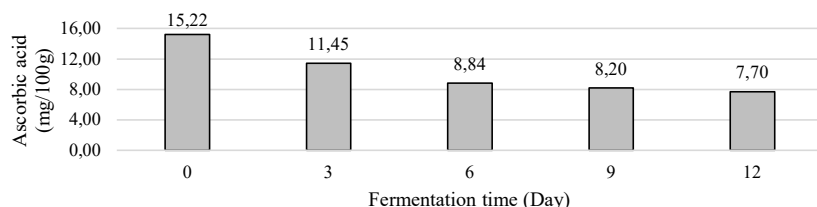
**Figure 1.** Changes in pH and reducing sugar content during rosella flower kombucha fermentation

Changes in OD<sub>600</sub> and dry weight of the mushroom (cellulose pellicle) during 12 days of roselle flower kombucha fermentation are depicted in Figure 2. The optical density (OD) at 600 nm, a measure of microbial growth, increases from 0.12 to 0.28, indicating a steady accumulation of microbial biomass. This finding is consistent with previous studies, such as those by Ahmed et al. (2020), which reported an increase in OD<sub>600</sub> corresponding with the growth of the kombucha SCOBY throughout fermentation. This microbial proliferation is correlated with a substantial increase in the mushroom dry weight, from 1.49 grams to 5.07 grams, with the most significant growth occurring after day 6. This growth pattern supports the findings of Charoenrak et al. (2023), who observed similar trends in cellulose pellicle development during kombucha fermentation. The term "mushroom" here refers to the cellulose pellicle produced by a symbiotic culture of bacteria and yeast (SCOBY), including species such as *Acetobacter xylinum*, known for its cellulose-synthesizing abilities. This suggests that the conditions created during the early stages of fermentation are favorable for microbial growth, which in turn supports the accumulation of mushroom biomass, likely through the activities of these bacteria and yeast in breaking down substrates that facilitate pellicle development.



**Figure 2.** Changes in optical density (OD) and mushroom dry weight during rosella flower kombucha fermentation

In relation to the ascorbic acid content, Figure 3 shows a marked decline from 15.22 mg/100g to 7.70 mg/100g over the fermentation period. The most rapid reduction occurs in the early stages (days 0 to 3), after which the decline slows and approaches a more stable value by day 12. This reduction in ascorbic acid could be attributed to its degradation under acidic conditions or microbial utilization. Vitamin C, an essential vitamin but unstable molecule, is prone to degradation due to its sensitivity to light, alkaline pH, high temperatures, oxygen, heavy metals, UV rays, and extended storage times. During fermentation, this degradation is exacerbated by the acidic conditions and the microbial activities involved. Specifically, the oxidation process and the presence of ascorbate oxidase enzymes produced by the fermenting microorganisms contribute significantly to the loss of ascorbic acid (Jafarpour & Hashemi, 2023).



**Figure 3.** Ascorbic acid content over time during rosella flower kombucha fermentation

The comparison between Roselle flower infusion and Roselle kombucha reveals several notable differences in their chemical and antioxidant properties (Table 1.). Roselle kombucha has a higher total phenol content (0.02  $\mu\text{L/mL}$ ) compared to Roselle flower infusion (0.01  $\mu\text{L/mL}$ ), suggesting that kombucha may offer a more substantial antioxidant potential due to its increased concentration of phenolic compounds. This is further supported by the higher % inhibition of DPPH at 25% (v/v) observed in Roselle kombucha (62.5%) compared to the Roselle flower infusion (55.88%), indicating that kombucha is more effective at scavenging free radicals. Additionally, the  $\text{IC}_{50}$  value for antioxidant activity, which reflects the concentration required to inhibit 50% of free radicals, is lower in Roselle kombucha (16.25%) than in Roselle flower infusion (16.85%), demonstrating that kombucha is slightly more effective as an antioxidant. Furthermore, when assessing lipase inhibition, Roselle kombucha exhibits a lower  $\text{IC}_{50}$  (0.147%) than Roselle flower infusion (0.253%), suggesting that kombucha is more effective in inhibiting lipase activity. The results of this study are in line with butterfly pea flower kombucha which is more effective in inhibiting pancreatic lipase in vitro than butterfly pea flower tea infusion (Aji et al., 2023b). These findings also align with previous research

indicating that kombucha, often fermented with various herbs and fruits, can have enhanced antioxidant and enzyme-inhibiting properties compared to its non-fermented counterparts. Studies such as those by Cardoso et al. (2020) and Jayabalan and Waisundara (2019) have shown that kombucha fermentation can increase the concentration of bioactive compounds and improve antioxidant capacities.

Table 1. Comparison of total phenol content, antioxidant activity, and in vitro lipase inhibition between roselle flower infusion and roselle flower kombucha

	Roselle flower infusion	Roselle flower kombucha (12 <sup>th</sup> day fermentation)
Total phenol content (GAE $\mu$ L/mL)	0,01	0,02
% inhibition of DPPH at 25% (v/v)	55,88	62,5
IC <sub>50</sub> antioxidant activity (% v/v)	16,85	16,25
IC <sub>50</sub> inhibition of lipase activity (% v/v)	0,253	0,147

The study comparing Roselle flower infusion and Roselle kombucha highlights several significant implications for health and functional beverage development. Roselle kombucha's superior antioxidant properties, evidenced by its higher total phenol content and better DPPH inhibition, suggest it could offer more effective protection against oxidative stress compared to Roselle flower infusion. This enhanced antioxidant activity may contribute to reduced risk of chronic diseases and support overall health. Additionally, kombucha's greater lipase inhibition suggests potential benefits for managing fat digestion and absorption, making it a promising option for weight management and metabolic health.

#### 4. Conclusion

The study demonstrates that rosella flower kombucha possesses superior antioxidant and in vitro lipase inhibitory properties compared to rosella flower infusion, likely due to the fermentation process that increases the concentration of bioactive compounds. The findings suggest that kombucha, particularly when fermented with functional ingredients like rosella, could serve as a potent health-promoting beverage. The enhanced antioxidant activity and lipase inhibition indicate potential benefits in managing oxidative stress and fat metabolism, making rosella kombucha a promising candidate for functional food development.

#### REFERENCES

- Agbor, G. A., Vinson, J. A., & Donnelly, P. E. (2014). Folin-Ciocalteu reagent for polyphenolic assay. *International Journal of Food Science, Nutrition and Dietetics (IJFS)*, 3, 1-10. <https://doi.org/10.19070/2326-3350-1400028>
- Ahmed, S., Mohamed, S., Hikal, M., Khadiga, A., & Abou-Taleb, M. (2020). Biological, chemical and antioxidant activities of different types Kombucha. *Annals of Agricultural Sciences*, 65, 35-44. <https://doi.org/10.1016/j.aos.2020.04.001>
- Aji, O. R., Rizqi, S. A., & Putri, D. A. (2023b). Antioxidant activity of Butterfly Pea Flower Kombucha (*Clitoria ternatea*). *Symposium of Biology Education (Symbion)*, 3, 298. <https://doi.org/10.26555/symbion.11749>
- Aji, O. R., Putri, N. F., & Putri, D. A. (2023a). Pancreatic lipase inhibitory activity of butterfly pea flower (*Clitoria ternatea*) Kombucha. *Biogenesis Jurnal Ilmiah Biologi*, 11(2), 151-159. <https://doi.org/10.24252/bio.v11i2.35724>
- Ariani, L. W., Ungsari, R. E. P., & Mighfar, S. (2019). Formulasi mikropartikel ekstrak etanol bunga rosela (*Hibiscus Sabdariffa* L.) dalam sediaan sirup dan potensinya sebagai obat herbal antiobesitas. *Jurnal Ilmiah Cendekia Eksakta*, 4, 1.
- Cardoso, R. R., Neto, R. O., D'Almeida, C. T. D. S., Nascimento, T. P. D., Pressete, C. G., Azevedo, L., Martino, H. S. D., Cameron, L. C., Ferreira, M. S. L., & De Barros, F. A. R. (2020). Kombuchas from green and black teas have different phenolic profiles, which impacts their antioxidant capacities, antibacterial and

**Commented [A3]:** Did authors conduct the assay in triplicate? If authors did so, please add standard error value as ... xx.xx  $\pm$  ee.ee

**Commented [A4]:** Did authors carry out statistical analysis? If authors did, the significant difference values can be added to the result, indicating significant decrease/increase in phenolics, etc.

**Commented [A5]:** Can authors explain why some values increase after fermentation?

**Commented [A6]:** Can authors suggest the optimal condition for roselle kombucha fermentation yielding optimal antioxidant and lipase inhibition activities?

antiproliferative activities. Food Research International, 128, 108782. <https://doi.org/10.1016/j.foodres.2019.108782>

Charoenrak, S., Charumancee, S., Sirisa-Ard, P., Bovonsombut, S., Kumdhithahutsawakul, L., Kiatkarun, S., Pathom-Aree, W., Chitov, T., & Bovonsombut, S. (2023). Nanobacterial cellulose from Kombucha fermentation as a potential protective carrier of *Lactobacillus plantarum* under simulated gastrointestinal tract conditions. *Polymers*, 15(6), 1356. <https://doi.org/10.3390/polym15061356>

Cohen, G., Sela, D. A., & Nolden, A. A. (2023). Sucrose concentration and fermentation temperature impact the sensory characteristics and liking of kombucha. *Foods*, 12(16), 3116. <https://doi.org/10.3390/foods12163116>

Gruenewald, J., Brendler, T., & Jaenicke, C. (2004). PDR for herbal medicines, third edition. Medical Economics Company, New Jersey.

Jafarpour, D., & Hashemi, S. M. B. (2023). Pure and co-fermentation of quinoa seeds by *Limosilactobacillus fermentum* and *Lactocaseibacillus rhamnosus*: Bioactive content, antidiabetic and antioxidant activities. *Fermentation*, 9(2), 80. <https://doi.org/10.3390/fermentation9020080>

Jayabalan, R., Malbaša, R. V., Lončar, E. S., Vitas, J. S., & Sathishkumar, M. (2014). A review on kombucha tea: Microbiology, composition, fermentation, beneficial effects, toxicity, and tea fungus. *Comprehensive Reviews in Food Science and Food Safety*, 13, 538-550. <https://doi.org/10.1111/1541-4337.12073>

Kim, D. H., Park, Y. H., Lee, J. S., Jeong, H. I., Lee, K. W., & Kang, T. H. (2020). Antiobesity effect of DKB-117 through the inhibition of pancreatic lipase and  $\alpha$ -amylase activity. *Nutrients*, 12, 3053. <https://doi.org/10.3390/nu12103053>

Kushargina, R., Rimbawan, R., Dewi, M., & Damayanthi, E. (2024). Effect of sugar addition and fermentation process on flavonoids, phenols, and antioxidant activity of Telang (*Clitoria ternatea* L.) kombucha. *Tropical Journal of Natural Product Research*, 8(3). <https://doi.org/10.26538/tjnpr/v8i3.39>

Naland, P. (2008). Kombucha teh dengan seribu khasiat. Agromedia Pustaka, Jakarta.

Nerdy, N. (2014). In silico docking of chemical compounds from roselle calyces (*Hibiscus sabdariffa* L.) as antidiabetic. *International Journal of ChemTech Research*, 7, 148-152.

Nopiyanti, D., & Harjanti, A. (2016). Analisis stabilitas senyawa aktif antioksidan kelopak bunga rosella (*Hibiscus sabdariffa* L.) pada penggunaannya sebagai bahan tambahan pangan alami. *Jurnal Farmasi Indonesia*, 13, 101-110.

Sari, P. (2014). Perbandingan aktivitas antioksidan kombucha teh hijau (*Camelia sinensis*) dengan teh daun mangga (*Mangifera indica*) dipengaruhi oleh lama fermentasi. Skripsi. Surakarta: Universitas Muhammadiyah Surakarta.

Susilowati, T. (2013). Perbedaan waktu fermentasi dalam pembuatan teh kombucha dari ekstrak teh hijau lokal Arraca kiara, Arraca yabukita, Pekoe dan Dewata sebagai minuman fungsional untuk antioksidan. Prosiding SNST ke-4. Semarang: Universitas Wahid Hasyim.

Wang, B., Rutherford-Markwick, K., Naren, N., Zhang, X., & Mutukumira, A. N. (2023). Microbiological and physico-chemical characteristics of black tea kombucha fermented with a New Zealand starter culture. *Foods*, 12(12), 2314. <https://doi.org/10.3390/foods12122314>

Windyaswari, A. (2018). Pengaruh teknik dan pelarut ekstraksi terhadap aktivitas antioksidan dan empat jenis ekstrak daun rosella (*Hibiscus Sabdariffa* L.). TM Conference Series, 1, 14-1. <https://doi.org/10.32734/tm.v1i3.254>

## 5. EMAIL PENGIRIMAN NASKAH REVISI

26 SEPTEMBER 2024



### Participants

Oktira Roka Aji (oktira)

Ichsan Luqmana Indra Putra (ichsanluqmana)

### Messages

Note	From
Please revise the manuscript as soon as possible according to reviewer comments	ichsanluqmana 2024-09-12 03:00 AM
<p>► Dear Editor,</p> <p>I hope this message finds you well. I am writing to submit the revised version of our manuscript for your consideration.</p> <p>We have addressed all the comments and suggestions provided by the reviewers and made the necessary revisions to improve the manuscript. A detailed response to each comment is included in the attached document.</p> <p>Thank you for the opportunity to revise our work. We look forward to your feedback.</p> <p>📎 <a href="#">oktira, Revisi JBNS 2024.docx</a></p>	oktira 2024-09-26 08:15 AM

Add Message

6. EMAIL KEPUTUSAN EDITOR: NASKAH DITERIMA

3 OKTOBER 2024

---

## [JBNS] Editor Decision

1 message

---

**Ichsan Luqmana Indra Putra** <ichsan.luqmana@bio.uad.ac.id>

Thu, Oct 3, 2024 at 9:07 AM

To: Oktira Roka Aji <oktira.aji@bio.uad.ac.id>, Anas Tasya Putri <anas1800017104@webmail.uad.ac.id>, Erisa Budi Handayani <erisa1800017135@webmail.uad.ac.id>, Nisa Ihsani <nisaihsani@umbandung.ac.id>

Oktira Roka Aji, Anas Tasya Putri, Erisa Budi Handayani, Nisa Ihsani:

*We have reached a decision regarding your submission to Journal of Biotechnology and Natural Science, "In Vitro Evaluation of Antioxidant and Lipase Inhibition Properties of Rosella (Hibiscus sabdariffa L.) Flower Kombucha".*

*Our decision is to: Accept Submission*

*We will notify you when your article is published.*

Ichsan Luqmana Indra Putra<br/>[ichsan.luqmana@bio.uad.ac.id](mailto:ichsan.luqmana@bio.uad.ac.id)



**C-Revisi JBNS 2024.docx**

85K



---

## [JBNS] Editor Decision

1 message

---

Eri <eri.eryati@staff.uad.ac.id> Fri, Oct 18, 2024 at 11:04 AM  
To: Oktira Roka Aji <oktira.aji@bio.uad.ac.id>, Anas Tasya Putri <anas1800017104@webmail.uad.ac.id>, Erisa Budi Handayani <erisa1800017135@webmail.uad.ac.id>, Nisa Ihsani <nisaihsani@umbandung.ac.id>

Oktira Roka Aji, Anas Tasya Putri, Erisa Budi Handayani, Nisa Ihsani:

The editing of your submission, "In Vitro Evaluation of Antioxidant and Lipase Inhibition Properties of Rosella (Hibiscus sabdariffa L.) Flower Kombucha," is complete. We are now sending it to production.

Submission URL: <http://journal2.uad.ac.id/index.php/JBNS/authorDashboard/submission/11271>